

PHYLOGENETIC POSITION AND TAXONOMIC DISPOSITION OF *TURRAEA BREVIFLORA* (MELIACEAE), A HITHERTO ENIGMATIC SPECIES

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SUMMARY

For decades, the rare Malesian shrublet hitherto called *Turraea breviflora*, was the only species of the pantropical family Meliaceae which could not confidently be placed in a genus. Previous morphological re-investigation led to the exclusion of the species from *Turraea* and suggested possibly close links with *Munronia*. In this study, parsimony, maximum likelihood and Bayesian analyses of DNA sequence data from plastid *rbcL* and nuclear ITS rDNA were used to infer the phylogenetic position of *T. breviflora* and its affinities to other representatives of the family. Phylogenetic analyses support Mabblerley's (1995a) proposal that the concept of the genus *Munronia* be expanded to accommodate it. The new combination *Munronia breviflora* (Ridl.) Mabb. & Muellner is therefore made.

Key words: Meliaceae, *Munronia*, Sapindales, *Turraea breviflora*, internal transcribed spacer (ITS), *rbcL*, taxonomy.

INTRODUCTION

Turraea breviflora Ridl. (Meliaceae; Fig. 1) is a rather inconspicuous suckering shrublet now restricted to Peninsular Malaysia. It seems that it was first collected by H.N. Ridley at Serangoon, Singapore in December 1897, but the plant has not been recorded from that island since; subsequently it was collected in Selangor and Johor in what is now Peninsular Malaysia.

When Ridley described the plant as a new species in 1922, he provisionally referred it to *Turraea* L., a genus otherwise unknown from the peninsula, or, indeed from the wetter parts of Malesia in general (Mabblerley 1995a, b). Since that time it has been collected on only a handful of other occasions and is still known from only a few localities in rather open hill Dipterocarp forest, often associated with bertam, *Eugeissona tristis*, and from limestone rocks. Paradoxically, it is known from some of the most well-worked sites such as Bukit Lagong and Ulu Gombak in Selangor, yet has never been found in fruit, and the number of flowering collections even from those sites is small. No doubt it is frequently overlooked, as it grows amongst tree seedlings and saplings which it greatly resembles. It is like no other Meliaceae and, in the field, sterile material may be readily confused with species of Icacinaceae or other families.

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Fig. 1. *Turraea breviflora* Ridl. a. Habit; b. half flower, with corolla removed. Drawing by Rosemary Wise, reproduced with permission from *Flora Malesiana Series I*, Vol. 12 / part 1 (1995) 28, f. 2.

Pennington & Styles (1975) noted that the relationships of the species were uncertain, and suggested it had “almost equal affinities with both *Turraea* and *Munronia*”. In the early 1980s, when preparing an account for *Flora Malesiana* Mabberley (1995a: 29), who had collected the plant in the field in 1974, examined herbarium material closely and made a critical morphological analysis, leading him to exclude *Turraea breviflora* from genus *Turraea* because of its habit, indumentum, corolla aestivation,

disk and the usual number of ovules in each locule. He pointed out that it resembled the genus *Munronia* Wight, as then understood, in its indumentum, but differed from that in its habit, valvate corolla, the usual number of ovules in each locule and possible dioecy. He thus concluded that it either represented an undescribed genus as Ridley had suspected, or that the concept of *Munronia* had to be enlarged. Today, it is the only species of the pantropical family Meliaceae which cannot confidently be placed in a genus. The writing by the second author of an account of the family Meliaceae for Flora of Peninsular Malaysia has prompted a re-examination of the plant, using molecular methods to augment the earlier morphological analysis, in an attempt to resolve its taxonomic placement.

In this study, therefore, we performed maximum parsimony (MP), maximum likelihood (ML) and Bayesian analyses of DNA sequence data from plastid *rbcL* and the internal transcribed spacers (ITS) of nuclear ribosomal DNA (nrDNA), defined as the unit containing the ITS1 spacer, 5.8S rRNA gene, and ITS2 spacer, to infer the phylogenetic position of *Turraea breviflora* and its affinities to other representatives of the family. Based on 46 species of Meliaceae, including a representative sampling for the two subfamilies, all tribes and almost all genera (44 out of c. 50), we focused on the following questions:

- What is the topological position of *T. breviflora* in the family phylogenetic trees derived from MP, ML and Bayesian analyses?
- What are the closest relatives of *T. breviflora*?
- Do the results support either of Mabberley's (1995a) taxonomic proposals?

MATERIALS AND METHODS

Plant material

Plant material was collected during field excursions, taken from herbarium specimens, or obtained from the DNA bank at Kew (<http://www.dnabank.kew.org>). *RbcL* sequences of 44 taxa of Meliaceae (ingroup) and 6 other taxa (outgroup) were available from the first author's previous work on Meliaceae (Muellner et al. 2003, 2006). Our *rbcL* matrix includes representatives of all tribes (Pennington & Styles 1975) of the only two currently recognised subfamilies (Muellner et al. 2003): Melioideae, viz. Turraeae, Melieae, Vavaeae, Trichilieae, Aglaieae, Guareeae, and Sandoriceae, and Cedreloideae (Swietenioideae), viz. Cedreleae, Swietenieae, Xylocarpeae, plus representatives of the Sapindalean families Sapindaceae, Rutaceae, and Simaroubaceae. ITS sequences of 32 taxa of Melioideae (ingroup) and four Cedreloideae (outgroup) were available from the first author's previous work on Meliaceae (Muellner et al. 2008a, b). Voucher information and GenBank accession numbers are listed in Table 1.

Isolation of DNA, amplification and sequencing

Total DNA from leaf fragments was extracted using a NucleoSpin Plant kit (Macherey-Nagel, Dueren, Germany). PCR amplification was carried out following Muellner et al. (2003, 2006, 2008a, b). PCR products were cleaned using a NucleoSpin® Extract II kit (Macherey-Nagel, Dueren, Germany). Sequencing reactions were run on a CEQ™ 8800 Genetic Analysis System (Beckman Coulter, Krefeld, Germany), following the manufacturer's protocol.

Table 1. Voucher information and GenBank accession numbers for samples used in this study (in alphabetical order). Voucher specimens are deposited in the following herbaria: ANBG = Australian National Botanic Gardens; FR = Herbarium Senckenbergianum, Senckenberg Research Institute; GC = University of Ghana; KEP FRI = Forestry Institute Malaysia; K = Royal Botanic Gardens, Kew; L = Nationaal Herbarium Nederland, Leiden University branch; LBC = Limbe Botanic Garden; NCU = University of North Carolina; WU = University of Vienna.

Taxon	Voucher (Herbarium)	GenBank accession number: <i>rbcL</i> , ITS.
<i>Aglaia elaeagnoidea</i> (A.Juss.) Benth.	<i>Samuel 4</i> (WU)	AY128209
<i>Aglaia odorata</i> Lour.	<i>Greger 903</i> (WU)	AY695552
<i>Ailanthus altissima</i> (Mill.) Swingle	<i>Chase 126</i> (NCU)	AY128247
<i>Allophylus cobbe</i> (L.) Raeusch.	<i>Chase 1306</i> (K)	AY128248
<i>Anthocarapa nitidula</i> (Benth.) T.D.Penn. ex Mabb.	<i>Chase 3313</i> (K)	DQ238056, DQ861616
<i>Aphanamixis polystachya</i> (Wall.) R.Parker	<i>Chase 2109</i> (K) <i>Samuel 14</i> (WU)	AY128213 AY695584
<i>Astrotrichilia</i> sp.	<i>Richard 25</i> (K)	DQ238060, DQ861621
<i>Azadirachta indica</i> A.Juss.	<i>Samuel 5</i> (WU)	AY128215, AY695594
<i>Cabralea canjerana</i> (Vell.) Mart.	<i>Pennington 17067</i> (K)	DQ238055, DQ861617
<i>Calodecarya crassifolia</i> Leroy	<i>Croat 31521</i> (K)	AY128216, DQ861631
<i>Capuronianthus mahafalensis</i> Leroy	<i>Fosberg 52439</i> (K)	AY128218
<i>Carapa guianensis</i> Aubl.	<i>Chase 2111</i> (K)	AY128219
<i>Cedrela odorata</i> L.	<i>Chase 2112</i> (K)	AY128220, DQ861606
<i>Chisocheton macrophyllus</i> King	<i>Chase 1309</i> (K)	AY128221, DQ861613
<i>Chukrasia tabularis</i> A.Juss.	<i>Chase 1308</i> (K)	AY128223
<i>Cipadessa baccifera</i> (Roth) Miq.	<i>Chase 1310</i> (K)	AY128224, DQ861627
<i>Dysoxylum gaudichaudianum</i> (A.Juss.) Miq.	<i>Chase 1312</i> (K)	AY128227, DQ861619
<i>Ekebergia capensis</i> Sparrm.	<i>MG 246</i> (Cynthia Morton)	AY128228, DQ861623
<i>Guarea glabra</i> Vahl	<i>Chase 336</i> (NCU)	AY128229, AY695591
<i>Harpullia arborea</i> (Blanco) Radlk.	<i>Chase 2018</i> (ANBG)	AY128249
<i>Heckeldora staudtii</i> (Harms) Staner	<i>Chase 3311</i> (K)	AY128230, AY695592
<i>Humbertioturraea</i> sp. (<i>H. labatii</i> Lescot ined.)	<i>Bardot-Vaucoulon 160</i> (K)	DQ238058, DQ861632
<i>Khaya anthotheca</i> C.DC.	<i>Chase 2859</i> (K)	AY128231, DQ861608
<i>Lansium domesticum</i> Corrêa	<i>Chase 2113</i> (K)	AY128232, AY695586
<i>Lepidotrachilia volkensis</i> (Gürke) J.-F.Leroy ex Styles & F.White	<i>Hughes 189</i> (K)	DQ238061, DQ861620
<i>Lovoa sywnnertonii</i> Baker f.	<i>Chase 2860</i> (K)	AY128233
<i>Malleastrum mandanense</i> Leroy	<i>Cheek et al. 3-17-5</i> (K)	DQ238062, DQ861626
<i>Melia azedarach</i> L.	<i>Chase 2867</i> (K)	AY128234, AY695595
<i>Munronia humilis</i> (Blanco) Harms	<i>Maxwell 75-239</i> (L)	EU621669, FJ194495/FJ194496
<i>M. pinnata</i> (Wall.) Theob.	<i>Samuel 6</i> (WU)	AY128236, DQ861604
<i>Naregamia alata</i> Wight & Arn.	<i>Kanodia 89603</i> (K)	DQ238059, DQ861629
<i>Nymanina capensis</i> (Thunb.) Lindb.	<i>Chase 270</i> (NCU)	AY128238, DQ861633
<i>Owenia vernicosa</i> F.Muell.	<i>Evans M3071</i>	DQ238063, DQ861622
<i>Pseudobersama mossambicensis</i> (Sim) Verdc.	<i>Bidgood, Abdallah & Vollesen 1426</i> (K)	DQ238064
<i>Pseudocedrela kotschyi</i> Harms	<i>Asase 008</i> (GC)	DQ238069/70
<i>Pseudoclausena chrysogyne</i> (Miq.) T.P.Clark	<i>Muellner 2052</i> (FR)	DQ238065, DQ861602
<i>Quassia amara</i> L.	<i>Fernando 1381</i> (LBC)	AY128250
<i>Quivisanthe papinae</i> Baill.	<i>Phillipson 1650</i> (K)	AY128239, DQ861605
<i>Reinwardtiodendron kinabaluense</i> (Kosterm.) Mabb.	<i>ALFB 112/87</i> (K)	DQ238054, AY695589
<i>Ruagea pubescens</i> H.Karst.	<i>Muellner 13</i> (WU) <i>Pennington & Frere 13761</i> (K)	DQ238057 AY695593
<i>Ruta graveolens</i> L.	<i>Chase 510</i> (K)	AY128251
<i>Sandoricum koetjape</i> (Burm.f.) Merr.	<i>Muellner 2050</i> (FR)	DQ861600, DQ238068

<i>Schmardaea microphylla</i> (Hook.) H.Karst. ex C. Muell.	Chase 746 (K)	AY128240
<i>Simarouba glauca</i> DC.	Chase 124 (NCU)	AY128252
<i>Swietenia macrophylla</i> King	Chase 250 (NCU)	AY128241, DQ861609
<i>Synoum glandulosum</i> (Sm.) A. Juss.	Chase 3314 (K)	AY128242
<i>S. glandulosum</i> (Sm.) A. Juss.	Schodde 5101 (K)	DQ861618
<i>Toona</i> sp.	Terrazas s.n. (K)	AY128243, DQ861607
<i>Trichilia emetica</i> Vahl	Chase 552 (K)	AY128244
	Sieglstetter 15 (FR)	EF136577
<i>Turraea breviflora</i> Ridl.	Kamarudin KEP FRI 51354 (KEP FRI)	EU621670, FJ194497
<i>Turraea heterophylla</i> Sm.	Küppers 2212 (FR)	EF136578
<i>Turraea sericea</i> Sm.	Civeyrel 1336 (K)	AY128245, DQ861630
<i>Vavaea amicorum</i> Benth.	Katik et al. 74722 (K)	DQ238066/67, DQ861610
<i>Walsura tubulata</i> Hiern	Chase 1314 (K)	AY128246, DQ861625
<i>Xylocarpus moluccensis</i> (Lam.) M. Roem.	Sandom 40 (K)	DQ238071

Sequence editing and alignment

Editing and assembly of the complementary strands were carried out with SeqMan™ II version 5.07 (Lasergene, DNASTAR, Inc., Madison, WI, USA). Alignment of *rbcl* sequences was performed by eye. Alignment of ITS sequences was performed following Muellner et al. (2008a, b). New sequences have been deposited in GenBank under the accession numbers EU621669–621670, FJ194495/FJ194496 and FJ194497 (<http://www.ncbi.nlm.nih.gov/>).

Phylogenetic analysis

MP analyses of the *rbcl* dataset were performed using PAUP* 4.0b10 (Swofford 2002). Substitutions at each nucleotide position were treated as independent, unordered, multi-state characters of equal weight (Fitch parsimony; Fitch 1971). Heuristic searches were carried out using 1000 random additions of taxa, tree bisection-reconnection (TBR) branch swapping, and the option MulTrees (keeping multiple, shortest trees), but holding only ten trees per replicate to reduce time spent in swapping on large numbers of trees. After 1000 replicates, we then used the shortest trees found as starting trees for a swapping-to-completion search (but with a tree limit of 10 000). Robustness of clades was estimated by bootstrapping (Felsenstein 1985) with 1000 replicates, using simple sequence addition, TBR branch swapping, and MulTrees, again holding ten trees per replicate. We consider 75–84% bootstrap values moderate support and 85–100% strong support. MP analyses of the ITS dataset and the combined *rbcl*/ITS dataset were conducted as described above, except for setting no constraint on the number of trees held per replicate during heuristic searches.

ML analyses were carried out with RAxML version 2.2.1 (Stamatakis 2006; <http://icwww.epfl.ch/~stamatak/index-Dateien/Page443.htm>) and PAUP* 4.0b10 (Swofford 2002), and Bayesian analyses were performed with MrBayes version 3.1.2 (Ronquist & Huelsenbeck 2003; <http://mrbayes.csit.fsu.edu/>). The substitution models employed in these analyses were found using Modeltest version 3.06 (Posada & Crandall 1998; <http://darwin.uvigo.es/software/modeltest.html>), which indicated the general time reversible model as best fitting our data with a proportion of invariable sites and a gamma

shape parameter alpha to model rate heterogeneity (GTR + I + G). For the Bayesian analyses, model parameters were estimated directly during two parallel runs, using four simultaneous chains and 2 000 000 cycles, sampling one tree every 100 generations. Trees that preceded stabilization of the likelihood value were excluded, and the remaining ones were used to calculate posterior probabilities via the construction of a majority rule consensus tree in PAUP. For the ML searches with RAxML we employed the GTR + G model, using 25 rate categories (instead of four as used in the Bayesian analyses).

RESULTS

The aligned *rbcL* matrix consisted of 1387 characters. For the *rbcL* matrix, 277 (20%) positions were variable and 166 (12%) were potentially parsimony informative. With the limits imposed as described above, the parsimony search produced 10 000 most parsimonious trees of 552 steps with a consistency index (CI) = 0.53 and a retention index (RI) = 0.81. The aligned ITS matrix consisted of 785 characters; 480 (61%) positions were variable and 366 (47%) were potentially parsimony informative. The parsimony search produced 10 most parsimonious trees of 2072 steps with CI = 0.41 and RI = 0.46. The combined *rbcL*/ITS matrix consisted of 2172 characters; 671 (31%) positions were variable and 464 (21%) were potentially parsimony informative. The parsimony search produced 2 most parsimonious trees of 2453 steps with CI = 0.41 and RI = 0.50.

Figures 2 and 3 show results of the ML and Bayesian analyses. *Turraea breviflora* is phylogenetically closest to the two representatives of the genus *Munronia* (Fig. 2, 3; also true for the single *rbcL* and ITS ML and Bayesian trees, trees not shown). Accessions of *T. breviflora* and two species of *Munronia* appear in a clade (Fig. 2, 3). The monophyly of the group is very weakly supported in the Bayesian analysis of the *rbcL* dataset (59% posterior probability, PP, tree not shown), and receives strong support in the combined *rbcL*/ITS analysis (100% PP, Fig. 3). Although *T. breviflora* and the two species of *Munronia* are monophyletic in the combined MP strict consensus tree, the clade does not receive > 50% bootstrap percentage; the latter is also true for the single *rbcL* and ITS analyses (trees not shown). In all three ML analyses, *T. breviflora* and *Munronia* appear in one clade (Fig. 2, and trees not shown).

DISCUSSION

Mabberley (1995a) argued, on morphological grounds, that, although *Turraea breviflora* belongs in Turraeae, it should be excluded from *Turraea* and is most closely related to *Munronia*; this study 1) confirms the position of *T. breviflora* in a clade comprising representatives of Turraeae and Trichilieae; 2) provides evidence that it is phylogenetically closest to *Munronia*; 3) suggests it indeed be referred to *Munronia*.

It should be noted here, however, that in order to reach a robust phylogenetic appreciation of Turraeae and Trichilieae, sampling of additional taxa on species level and the collection of additional DNA data will be necessary.

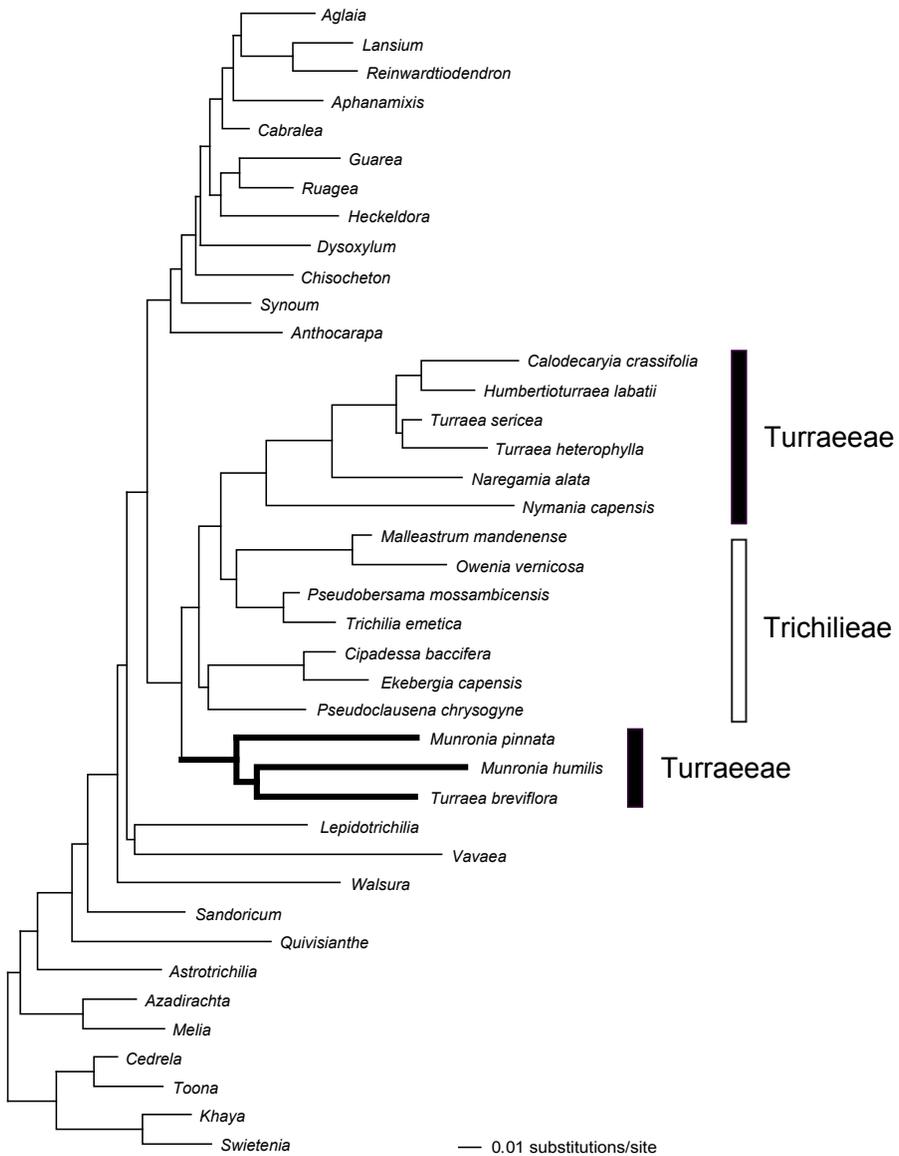


Fig. 2. ML tree obtained from the ML analysis of the combined *rbcL*/ITS dataset of 36 Melioideae ingroup and four Cedreloideae outgroup accessions. Subfamilies after Muellner et al. (2003).

Mabberley's (1995a) proposal that the concept of the genus *Munronia* be expanded, is vindicated by our new findings, so that an amended description of the genus is provided below.

The last enigmatic species of Meliaceae has found a home.

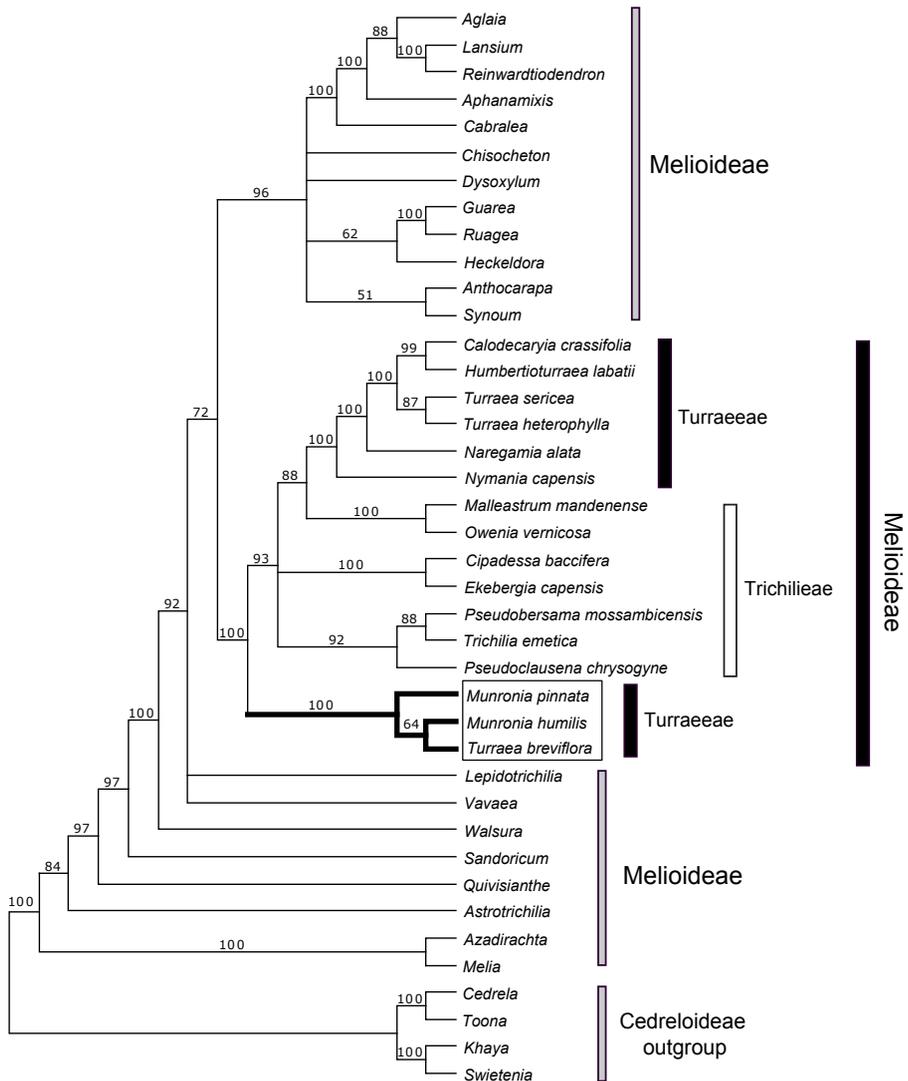


Fig. 3. Bayesian tree (10 000 total trees, burn-in of 1000 trees) of the combined *rbcL*/*ITS* dataset of 36 Melioideae ingroup and four Cedreloideae outgroup accessions (50% majority rule consensus tree). Subfamilies after Muellner et al. (2003). Numbers above branches are Bayesian posterior probabilities.

MUNRONIA

Munronia Wight (1838). — Type species: *Munronia pumila* Wight (= *M. pinnata* (Wall.) Theob.).

Unbranched or sparsely branched shrublets, sometimes suckering with apparently short-lived shoots, sometimes possibly dioecious. *Indumentum* a mixture of simple and bifid or stellate hairs. *Leaves* simple to imparipinnate, often crenate to serrate.

Selected specimens:

PENINSULAR MALAYSIA. **Selangor**: Kepong, Bukit Lagong Forest Reserve, above reservoir, *Mabberley 1566* (FHO, K, KEP) & 16th mi, Genting Simpah Road, Selangor Ridge, *Kochummen FRI 2971* (K). **Johor**: Sungei, Kau Ara, *Corner SFN 31470* (K).

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