A NATURAL INFRAGENERIC CLASSIFICATION FOR CICER (LEGUMINOSAE, CICEREAE)

A.M.R. DAVIES¹, N. MAXTED² & L.J.G. VAN DER MAESEN³

SUMMARY

A comprehensive morphological survey and analysis of all taxonomically recognised wild species of *Cicer* L. (Leguminosae, Cicereae) is presented. The data (104 characters from 152 herbarium specimens representing 34 of the 44 recognised taxa in the genus *Cicer* with supplementary data for the remaining taxa taken from the literature) were analysed using multivariate statistics (cluster analysis, factor analysis and ordination techniques). The results are discussed in the context of extant classifications and the re-organisation of a novel infrageneric classification also incorporating information from published genetic data. A revised classification with 3 subgenera, 5 sections and 2 series is proposed.

Key words: Leguminosae, Cicereae, infrageneric classification, morphology, wild chickpeas.

INTRODUCTION

The genus Cicer (Leguminosae, Cicereae) comprises 44 species (C. tragacanthoides has two varieties), 9 annuals and 35 perennials (Van der Maesen et al., 2007), two more than Coles et al. were able to report (1998), which have a centre of diversity in south-western Asia, with remote, endemic species found in Morocco and the Canary Islands (Van der Maesen, 1987). The genus is the only member of the tribe Cicereae Alef., subfamily Papilionoideae, family Leguminosae. It was historically included in the legume tribe Vicieae (Bronn) DC., but Kupicha (1977) presented detailed taxonomic evidence to support the tribal distinction of the genus from the other Vicieae genera: Vicia L., Pisum L., Lens Adans., Lathyrus L. and Vavilovia Fed. To this end Kupicha (1977) reinstated the monogeneric tribe Cicereae originally proposed by Alefeld (1859) and provided a detailed generic description (Kupicha, 1981). The most widely known species is the cultivated chickpea, Cicer arietinum L. Much attention has focussed on the cultivated form and its subspecific taxa in the past. However, in more recent years the focus of interest has broadened to include all the annual species because of the search for various beneficial crop-related traits in wild Cicer species (Muehlbauer et al., 1994; Hannan et al., 2000). A consequence of this research bias is that the perennial species, largely located in remote Central Asia, have been under-collected and taxonomically neglected until recently. After the genus was revised by Van der Maesen (1972), several

¹⁾ Institut für Systematische Botanik, Menzinger Strasse 67, München, Germany.

University of Birmingham, School of Biosciences, Edgbaston, Birmingham, B15 2TT, United Kingdom.

Nationaal Herbarium Nederland, Wageningen branch (Herbarium Vadense), Biosystematics Group, Wageningen University, Generaal Foulkesweg 37, 6703 BL Wageningen, The Netherlands.

	Of a section (1) as	unwhidouare			
			Subaphylum		
Mecroarthum		uthoidea	Macracanthum		
Acanthocer Tragecarthodea		Tragaca	Tragacanthoidea		
Pungens			Pungens		
Macrosoft um Macrosoft um			Anatolo- Perso- Orietalia		
Acanthocicer Tragacanthocidea Wolestum	Marked and	Viciastrum ficioides	Anatolo- Persica		
Pungeins Pungeins			Flexuosa		
Microphylle Microphylle Micros Pasisie Peisise	Cicer		Persica		
Acanthrocicer Songarica Clear Langueo-		ioer	Europae o- Anatolica		
American Ame		Polye	Graeca	Ctonophillium	Sterioprismum
ocer Analop Person Person Mano- Polycier Mano-		Nano- Polucion	sound in a	ľ	Virinidae
Victaethum March-Poly March-Poly Marchaether Austolica Austolica Perennia		Der	Perennia	bin dia an	Bolininar
Tragacanthotiea Coer Paroca Paroca Annua		Chamaeck	Annua	Cicer	Annua
Ceerr Arietina Apiculata Tragacanthoidea Paynocer Polycienr Polycienr Polycienr Polycienr	V	CIOR	Macro- aristae		Chamaarirar
Sprocenses Cumblen Comblen Commence Chamaeccer Damaeccer Monoccer		Monocioer	Cirthifera	Ciner	
Victories https/pmilta Arterna Parado Parado Parado Parado Arterna			Arietina		Si C
G Sted. G G G G G G G Stab G Stab G Stab G Stab G Stab G Stab G Stab G Stab G Stab G Stab G Stab G Stab G G G G G G G G G G G G G G G G G G G	9	Sect.	Ser.	9.90	Sort S
Jundent & Spacch		Seferova (1995)	~		Davies & al. (2008)

Fig. 1. Taxonomic history of the infrageneric hierarchies in Cicer L.

new *Cicer* species have been described. The current exploitation interest and addition of new taxa have resulted in the need for a revised classification of the genus. It has been increasingly clear that the traditionally defined infrageneric groupings result in non-monophyletic species, subgenera and sections (e.g., Rajesh et al., 2003; Frediani & Caputo, 2005). The general aim of this research was to clarify the relationships between *Cicer* taxa by combining multivariate analytical approaches with morphological data and the published genetic data and produce a revised natural classification that will aid future collection, conservation and utilisation of the genus. Interactive keys and short descriptions have been published by Van der Maesen et al. (2007), previous to the present paper of which publication was inadvertently delayed.

The genus *Cicer* was first studied in detail by Jaubert & Spach (1842), who described four sections, *Arietina, Vicioides, Spiroceras* and *Tragacanthoides* based on woodiness and terminal structure of the rachis (presence of a leaflet, spine or tendril). The infrageneric classification of *Cicer* over the last 165 years is illustrated in Fig. 1. Both formal and informal (Alefeld, 1859) hierarchies are included, as are the nomenclatural invalid series of Linczevski (1948). Boissier (1872) adopted this system but merged sections *Vicioides* and *Spiroceras*. Popov (1929) proposed a more detailed classification that subdivided the genus into 2 subgenera and 4 sections, also incorporating subsectional, series and subseries taxa. Linczevski (1948) revised Popov's classification streamlining the divisions within the sections.

The genus was comprehensively revised by Van der Maesen (1972), who extended the work of Popov (1929) and Linczevski (1948). The classification proposed by Van der Maesen contained 2 subgenera, *Pseudononis* Popov and *Viciastrum* Popov (distinguished by flower size and calyx morphology) and 4 sections distinguished as follows: *Monocicer*, annual, small-flowered species with firm, erect to inclined or prostrate stems and imparipinnate leaves, or the rachis terminating in a tendril; *Chamaecicer*, annual or perennial shrubby species with thin creeping branches and 3–7 leaflets per leaf; *Polycicer*, the large-flowered perennial species with imparipinnate leaves or tendrilous rachis; and *Acanthocicer*, large-flowered species with persistent spiny rachis and spinous calyx teeth. The sections are further subdivided into 2 subsections and 14 series. The characters used to divide the series included: standard length, number of pairs of leaflets on the rachis, apical structure of the rachis, number of flowers per inflorescence, bract and stipule characters, arista size and life cycle.

The *Cicer* taxa present within the former Soviet Union were reviewed by Seferova in 1995. As well as lectotypifying the supraspecific ranks, she replaced some illegitimate names, effectively publishing others and drew attention to several new Central Asian species. The amended classification includes 3 subgenera, 8 sections and 13 series. The characters she used to distinguish taxa are primarily the ones used previously by Van der Maesen (1972). The most significant difference between the classifications of Van der Maesen (1972) and Seferova (1995) is seen in the sections of subgenus *Cicer*, and sections *Polycicer* Popov, *Nanopolycicer* (Popov) Seferova and *Vicioides* Jaub. & Spach of subgenus *Viciastrum*, and in the series divisions of section *Tragacanthoidea* Jaub. & Spach.

Both the Van der Maesen (1972) and Seferova (1995) classifications were produced using non-numeric techniques. Seferova proposed several nomenclatural changes to the taxa accepted by Van der Maesen (1972), reducing several entities to synonyms as follows: *C. macracanthum* Popov (= *C. incanum* Korotkova); *C. grande* (Popov) Korotkova (= *C. paucijugum* Nevski); *C. macracanthum* subsp. *acanthophyllum* (Boriss.) Seferova (= *C. acanthophyllum* Boriss. = *C. garanicum* Boriss.); *C. flexuosum* subsp. *mogoltavicum* Popov (= *C. mogoltavicum* Popov). In addition, several new *Cicer* species have been described since 1972: *C. canariense* A.G. Guerra & G.P. Lewis (Santos Guerra & Lewis, 1985); *C. heterophyllum* Contandr., Pamukç. & Quézel (Contandriopoulos et al., 1972); *C. reticulatum* Ladiz. (Ladizinsky, 1975); *C. luteum* Rassulova & Sharipova (Rassulova & Sharipova, 1992); *C. laetum* Rassulova & Sharipova (Rassulova & Sharipova, 1978) and *C. rassuloviae* Lincz. (Linczevski, 1948). The infrageneric structure of *Cicer* has become progressively more unwieldy over time. The complex hierarchy attempts to justify the perceived interspecific variation based on species whose infraspecific variation is often poorly understood.

New information from biochemistry, plant breeding and especially genetics have highlighted the weaknesses in the current infrageneric hierarchy, indicating that the existing perception of species relationships based on the congruence of a few characters cannot adequately explain true natural relationships in the genus.

Therefore, to clarify the relationships between taxa within the genus *Cicer* it is time to update and re-evaluate the infrageneric classification using objective techniques.

MATERIALS AND METHODS

Forty-one species and four varieties were included in the analysis. The characteristics of thirty-three species were recorded from 152 herbarium specimens and the remaining taxa were scored from literature sources. Herbarium specimens were loaned from BM, E, K and WAG (abbreviations follow Holmgren et al., 1990). Author names follow Brummitt & Powell (1992). A species citation list is given in the Identification List at the end of this article. The character set was compiled from the literature (Davis, 1970; Contandriopoulos et al., 1972; Van der Maesen, 1972, 1987; Kupicha, 1977; Coles, 1993; Maxted, 1993) and personal observations of the specimens. The data matrix is provided in Table 1 and the characters and character states recognised are listed in Table 2. The final character list contained 104 characters, including: 14 habit and stem characters, 36 vegetative characters, 25 inflorescence characters, 16 pollen, anthers, pods and seeds characters and 13 characters covering features such as phenology, altitude and chromosome number. Many different types of character were recorded. Continuous data, usually considered to be intrinsically ordered when coded into 'discrete' states, were scored as numbered states accounting for the natural ranges of variation. E.g. rachis length (character 19; Table 2) was seen to vary in three natural clusters throughout the data set: those species whose rachis was persistently less than 40 mm long, those with rachis' between 40 and 70 mm long and those with long rachis' over 70 mm. All characters were scored as either binary or multistate but considered as unordered in the final analysis. The code of 9 was used to represent missing data. Where appropriate during the analyses non-variable characters, and characters for which there was a lot of missing data (e.g. characters 86-90, 93-104), were omitted.

Within *Cicer* there have been several non-morphologically-based investigations of species relationships, particularly among the annual species. Studies have covered molecular variation (Kazan & Muehlbauer, 1991; Ahmad & Slinkard, 1992; Ahmad et al.,

Table 1. Data matrix. The species names are indicated by the first four letters of the specific epithet, or in the case of varieties or subspecies the first letter of the specific epithet and the first 3 letters of the infraspecific epithet. There are 43 taxa and 106 characters, the codes of which can be found in Table 2. '9' denotes

missing	values.										
acan	0011019211	0221091100	0101120102	0021011100	0100102101	0111011100	3310010121	1222192001	1219921011	1199999999	6666
anat	0111020111	2221010111	0010100101	1321101000	0212111122	1211001901	3320110120	2202119011	0012011011	1199299909	9993
arie	1100020222	1211000111	2011020211	1121110100	0220001122	1201001111	1000100011	0001113010	0009901101	0100000010	0233
atla	1010209200	0210000100	2911020012	0101001111	1100191011	1200001111	3200010199	6006000066	1219920010	1199999999	6666
balc	1010021132	0119990121	0910010100	2921100111	0999091221	1121009911	9320111999	9902111001	0999910011	6666666666	6666
bald	0010020020	0229990122	0110010110	2331011011	0999011120	1121001111	4920001220	1009909999	9909910011	1299999999	6666
biju	0000123100	0020010001	2111020101	1121101011	0222121111	1231001110	3201011111	0001011111	1112001211	0111111012	2011
cana	1010110222	0112210320	2010100119	2011210100	1000111191	1000002111	2310001021	1002010110	1212200322	029999999	9992
chor	0000012200	212000002	2211020012	0200002111	1000111001	1100001110	2000010091	0010020120	1200111012	0091222929	1666
cune	1001111210	0212210111	2010010111	1021000100	0120111122	1200001100	3200000011	0001119210	0201101221	0123333939	9021
echi	1000222210	2020100101	2011020100	1121201100	0120121121	1221201111	3300000011	0210123109	1110301201	010000100	2291
fedt	0111021100	0110010211	0010000101	1221111100	0900011121	0210201101	1320010221	2020191001	1202021011	1199999999	6666
flex	0011021221	1111210111	0210010112	1221111100	0102011110	1111011101	4320101221	2210119011	1202001011	1299999999	6666
flor	1011003211	01111001111	2110000010	2321111000	0220021231	1011002101	3310011991	9200119211	1211100311	1299999999	0666
grae	1011012220	2110010111	2010010010	1241111000	0222011231	1031102101	3310010121	2390011211	0002010311	1299999999	6666
gran	0111011122	0219990222	0110010110	2321111001	0202021220	1121001111	0920110120	2202193000	1209910011	1199999999	6666
hete	1010000932	0119921129	9911020910	2221209999	9999991092	1099299999	9199999999	6666666666	9999900311	6666666666	1666
inca	0119003910	0129991101	9901120999	0201001111	1999191122	1119099111	1920101991	1299999999	9909920010	1199999999	6666
inci	0011210200	101000000	2011020011	0121102101	1120101121	0201001110	1310000011	0209011011	0300111010	66666666610	1666
isau	0011020121	0111200112	2011020000	2321121111	1200021111	1021001101	1020001121	1409199909	9201000012	1299999999	1666
juda	1000120201	2020100100	2111020100	0121100110	1122101021	1200002110	3201010002	0110011120	1200001201	0111111110	1121
kerm	1111120132	0211100221	0010100012	0100011110	00001110001	1221001110	9021000121	1391111001	1001020011	66666666610	6666
kors	0111011132	9229990111	0100000100	2321111100	0299011230	0229009911	19999999999	9902919999	9909911011	6666666666	6666
macr	0111020121	0021001201	0101120012	0131011100	0100112211	0211011101	3320000021	1021111011	1210121011	1199999999	6666
micr	0011121110	1211110221	0010000112	1221001100	0100021120	1121001101	1020010121	2002110010	0002021011	1199999999	0666
mogo	0111020230	0219990222	0110110112	0200011101	0199101122	1121001111	3320010220	1002112010	1209900011	1299999999	6666
mont	1010011120	0112290111	2111020110	2341121101	1200021230	1021211001	3011011121	1302111021	2212001311	1299999999	9993
mult	0011121110	0011120221	0011020101	1121111000	0199011221	1121001101	2320000121	2029190021	1202030011	1199999999	6666
nuri	0011022120	0112000122	0010010010	1321110000	0299021120	1221011101	0320111221	2099112001	0212021319	1299999999	6666
oxyo	1110020222	1111200122	0010110000	1330111110	0202021021	1111011011	2010001120	1012112010	1210111010	1299999999	6666
pauc	0111021111	0029990001	2101020110	1221101111	0199101130	1211001101	0320011120	1001999999	9909921011	1299999999	6666
pana	1000121210	2210000101	2011020101	0121100111	1120211021	1201001110	3201010000	0011110020	1200101210	0111111112	1121
ppin	1000122200	2220000100	2011020101	0021101111	1100211122	1201001110	3201010000	0010010020	1200301212	0111111112	1121
gund	0111023211	0110001101	0201120011	1200011100	0000012111	1211001101	0310101121	1111012011	1200121011	1199999999	0666
rech	0111120121	0111201210	0201120112	0101111100	0000001111	1111001101	0310000121	0192019999	9910020011	1199999999	6666
reti	1000021212	0010000100	2111020211	1141111100	0100011190	1201001110	0310099999	9911013010	1099990911	0100000010	0201
song	0111021122	0101120211	0210000112	1220011100	0100021130	0111202101	2320110221	2022013000	2212011011	1299299999	6666
spir	0111020222	0112100122	0010100012	0230011110	0102022020	1111001111	3311010120	1512112011	1211010012	029999999	6666
stap	0111020210	0119991101	2101121011	1030101100	0003002101	1110001111	3921110021	1121112001	1009930011	1199999999	6666
suba	0111029311	0112031122	2001121119	0011221100	0033002111	01110011101	3310001010	0011112001	1210010011	02999999999	6666
trag	0111021200	01110011101	0201120112	0000001100	0000002021	1211001100	1320110121	1021112010	0200021010	11 99999999	6666
ttur	0011021221	0111101111	0201120112	0100011100	0000002021	1211000101	1320110121	1012119099	9900021010	11 99 99 99 99	6666
yama	1100012201	2020100100	2911020011	2141101101	1120011011	1000011110	3200010001	0110021000	1211200011	0192229222	1191

Woodiness	0 = woody; 1 = not woody
Stem robustness	0 = slender; $1 =$ sturdy
Growth cycle	0 = annual; $1 = $ perennial
Habit	0 = herb; 1 = shrub; 2 = climber
Form	0 = erect; 1 = semi-erect; 2 = procumbent
Branching	0 = simple; $1 = $ sparse; $2 = $ secondary
Stem hair density (hairs/mm ²)	$0 = \langle 20; 1 = 20 - 30; 2 = 31 - 40; 3 = \rangle 40$
Stem hair type	0 = eglandular; $1 = $ glandular; $2 = $ both; $3 = $ glabrous
Stem height (mean cm)	$0 = \langle 20; 1 = 20 - 30; 2 = 31 - 40; 3 = \rangle 40$
Stem height (range cm)	0 = < 10; 1 = 10 - 20; 2 = > 20
Pigmentation	0 = absent; 1 = sometimes; 2 = always
Stem orientation	0 = straight; $1 = $ flexuous; $2 = $ both
Stem surface ribbed	0 = prominently; 1 = ribbed; 2 = faintly
Internodal length (mean mm)	$0 = \langle 20; 1 = 20 - 30; 2 = \rangle 30$
Internodal length (range mm)	0 = < 10; 1 = 10 - 20; 2 = > 20
Leaf hair length (mm)	0 = < 0.5; 1 = 0.5 - 0.89; 2 = > 0.9, 3 = no hairs
Leaf arrangement	0 = imparipinnate; 1 = paripinnate
Number of pairs of leaflets	0 = 1-4; 1 = 3-10; 2 = 7-15; 3 = > 20
Rachis length (mm)	0 = < 40; 1 = 40-70; 2 = > 70
Leaf spacing on rachis	
(character 18, 19)	$0 = 1-5; 1 = 6-11; 2 = \ge 12$
Leaf shape outline	0 = decrescent; $1 = $ increscent; $2 = $ equal
Rachis rolled	0 = loosely; $1 = $ tightly; $2 = $ not
Rachis apex – spine	0 = present; $1 = $ absent
Rachis apex – tendril	0 = present; $1 = $ absent
Rachis apex – leaflet	0 = present; $1 = $ absent
Tendril structure	0 = simple; $1 =$ branched; $2 =$ none
Leaflet form	0 = laminate; $1 = $ spinose
Phyllotaxy	$0 = opposite; 1 = \pm opposite; 2 = alternate$
Leaflet margins	0 = doubly incised; $1 =$ incised
Leaflet margin serrations	0 = entire -2/3; 1 = 1/2 - 2/3; 2 = apex -1/2
Leaflet length (mm)	0 = <5; 1 = 5-10; 2 = >10
Leaflet width (mm)	0 = 0-2; 1 = 2-4; 2 = 4-6; 3 = > 6
Leaflet shape	0 = cuneiform; $1 = $ linear; $2 = $ obovate; $3 = $ rotund; $4 = $ elliptic
Leaflets flabellate	0 = yes; 1 = no
Leaflet apex	0 = truncate; $1 =$ rounded; $2 =$ acute
Leaflet base	0 = cuneate; $1 = $ rounded-cuneate; $2 = $ truncate
Leaflet teeth shape	0 = acute; 1 = acuminate/triangular; 2 = rectangular/cuspidate
Terminal apical spine	0 = tendrillous; $1 = $ inflexed
Midrib tooth	0 = prominent; 1 = not
Midrib tooth	0 = spinose; $1 = $ not
Midrib tooth	0 = recurved; $1 = $ not
Number of teeth per leaflet	$0 = 0-5; 1 = 6-11; 2 = \ge 12$
Leaflet surface ribbed	0 = U = L; 1 = U > L; 2 = U < L; 3 = smooth
Leaflet surface pubescent	0 = U = L; 1 = U > L; 2 = U < L; 3 = glabrous
Leaflet teeth apices	0 = spine; $1 = $ mucro; $2 = $ none
Leaflet petioles	0 = sessile (0-0.3); 1 = subsessile (0.4-0.6);
*	2 = petiolate (0.7-1.0)
Stipules	0 = spiny; $1 = $ laminate; $2 = $ both
Stipule length (mm)	0 = 0.0 - 2.9; 1 = 3.0 - 5.9; 2 = > 6.0
Nr. teeth on stipules	0 = 1 only; $1 = 1-3$; $2 = 2-6$; $3 = > 6$

Table 2. Description of characters and their states.

50 Stipule outline 0 = flabellate/oblique; 1 = ovate/lanceolate; 2 = triangular 51 Stipule (basal) v. leaflet size 0 = equal or bigger; 1 = smaller52 Number of flowers / raceme 0 = some many flowered; 1 = none more than 2-flowered; 2 =always 1-flowered 0 = 0 - 15; 1 = 16 - 30; 2 = 31 - 45; 3 = > 4653 Peduncle length (mm) 54 Pedicel length (mm) $0 = \le 5; 1 = \ge 6$ 55 Arista form 0 = spine; 1 = leaflet; 2 = both

(Table 2 continued)

56	Arista length (mm)	$0 = 0-10; 1 = 11-20; 2 = \ge 21$
57	Bract shape	0 = spinose; $1 =$ triangular toothed; $2 =$ flabellate
58	Calyx teeth	0 = recurved; $1 = $ straight
59	Calyx dorsally gibbous	0 = strong; 1 = weak; 2 = not
60	Calyx length (mm)	0 = <7; 1 = >7
61	Calyx tooth shape	0 = triangular; 1 = broadly lanceolate; 2 = narrowly lanceolate;
	v 1	3 = lanceolate acuminate; $4 = $ cuspidate
62	Corolla colour	0 = white; $1 =$ yellow; $2 =$ pink; $3 =$ blue; $4 =$ purple
63	Corolla length (mm)	0 = 5 - 10; 1 = 11 - 15; 2 = 16 - 27
64	Corolla shape	0 = obovate; 1 = ovate
65	Corolla pubescence	0 = pubescent: $1 = $ glabrous
66	Corolla apex	0 = marginate; $1 = emarginate$
67	Corolla apex	0 = mucronate: 1 = not
68	Corolla width (mm)	0 = 4 - 9: $1 = 10 - 15$: $2 = > 16$
69	Filament length (mm)	0 = < 5; 1 = 6 - 10; 2 = > 11
70	Alae auriculate	$0 = \log \log(1 - 1)$ $1 = 10$ 10 , $2 = 2$ 11
71	Alse length (mm)	0 = < 10; 1 = 10 - 15; 2 = > 15
72	Alae shape	$0 = 0$ of $1 = 10^{-13}$, $2 = 215$ $0 = 0$ of $1 = 0$ of 10^{-13} , $2 = 215$
12	Mae shape	5 = triangular
73	Pod shape	0 = oblong; 1 = ovate; 2 = oblong; 1
74	Pod longth (mm)	0 = small(0, 10); 1 = madium(10, 20); 2 = large(5, 20)
74	Pod width (mm)	$0 = \sinh(0-10), 1 = \sinh(0-20), 2 = \arg(220)$
76	Pod width (illin)	0 = < 0, 1 = > 0 0 = color dylor 1 = clor dylor 2 = both 2 = clobroys
70	Pod hair type	0 = egrandular, 1 = grandular, 2 = bour, 5 = grabrous
70	Seed length mean (mm)	0 = 3; 1 = 4; 2 = 3; 3 = 0
70	Seeds beaked	0 = yes; 1 = no
/9	Seed snape	0 = obovate; 1 = globular; 2 = angular
80		0 = prominent; 1 = not
81	Seed coat colour	0 = black; 1 = brown; 2 = purple
82	Seed coat surface	0 = wrinkled; $1 = $ echinate; $2 = $ tuberculate; $3 = $ smooth
83	Calyx teeth	0 = equal; 1 = unequal
84	Calyx hair length (mm)	$0 = 0.0 - 0.5; 1 = 0.6 - 0.9; 2 = \ge 1.0$
85	Calyx hair density (hairs/mm ²)	$0 = \langle 20; 1 = 20 - 30; 2 = 31 - 40; 3 = \rangle 40$
86	Altitudinal range (m a.s.l.)	0 = < 1000; 1 = c. 2000; 2 = c. 3000; 3 = c. 4000
87	Endemic	0 = yes; 1 = no
88	Habitat	0 = rubble; $1 = $ cultivation; $2 = $ weedy; $3 = $ forest
89	Phenology	0 = JanApril; 1 = May-August; 2 = SeptDec.
90	Soil type – Geology	0 = Basic, calcicole; $1 = indifferent$; $2 = Acid$, volcanic
91	Calyx tube length (mm)	0 = 0.0 - 2.9; 1 = 3.0 - 5.9; 2 = 6.0 - 8.9; 3 = > 9.0
92	Calyx teeth length (mm)	0 = 0.0 - 2.9; 1 = 3.0 - 5.9; 2 = 6.0 - 8.9; 3 = > 9.0
93	Crossability (Ladizinsky &	
	Adler, 1976)	0 = I; 1 = II; 2 = III
94	Seed storage proteins (Ahmad &	
	Slinkard, 1992)	0 = I; 1 = II; 2 = III; 3 = IV
95	Isozyme variation (Tayyar &	
	Waines, 1996)	0 = I; 1 = II; 2 = III; 3 = IV
96	Isozyme polymorphism	
	(Ahmad et al., 1992)	0 = I; 1 = II; 2 = III; 3 = IV
97	Seed proteins (Sammour, 1994)	0 = I; 1 = II; 2 = III; 3 = IV
98	Disease resistance (Robertson	
	et al., 1996)	0 = greatest (> 5/7); 1 = average (3B4/7); 2 = least (0/7)
99	Allozyme variation and phylogeny	
	(Kazan & Muehlbauer, 1991)	0 = I: 1 = II: 2 = III: 3 = IV
100	Pollen morphology	,,,
	(De Leonardis et al., 1994)	0 = primitive: 1 = middle: 2 = derived
101	Seed coat (De Leonardis et al 1994)	0 = primitive: 1 = middle: 2 = derived
102	Pollen morphology (Ocampo 1997)	0 = primitive: 1 = middle: 2 = derived
103	Karvoptype morphology	- prima e, r = made, z = derived
100	(Kabir & Singh 1990)	0 = 7 pairs median: $1 = 5$ pairs median: $2 = 1$ subterminal
104	Chromosome numbers	• Paris median, 1 – 5 paris median, 2 – 1 subterninar
101	(Van der Maesen, 1987)	0 = 14; $1 = 16$; $2 = 24$; $3 = variable$

1992, Sammour, 1994; Tayyar & Waines, 1996; Javadi, 2004; Javadi & Yamaguchi 2004a, b; Frediani & Caputo, 2005, Javadi et al., 2007), seed coat and plumule morphology (Javadi & Yamaguchi, 2004c), palynology (Ocampo et al., 1992; De Leonardis et al., 1994), cytology (Van der Maesen, 1987; Kabir & Singh, 1990), crossability groups (Ladizinsky & Adler, 1976) and disease resistance (Robertson et al., 1996). However, these studies have rarely attempted a genus-wide examination and have generally focussed on the annual species closely related to cultivated chickpeas. Where appropriate, their results are included in the taximetric analysis and in some cases they have been central in drawing overall conclusions for interspecific relationships.

The data set was initially analysed using the program NTSYS (Rohlf, 1995) – Cluster Analysis. The program used Sneath's Simple Matching Coefficient to calculate an Euclidean distance based similarity matrix and this was analysed using the UPGMA clustering method. Cophenetic correlation can be calculated and used as an indication of degree of fit between the similarity matrix and the cophenetic value matrix based on the UPGMA cluster file. Rohlf (1995) defines the goodness of fit as a correlation coefficient *r*, based on the Mantel statistic *Z*. A poor fit is deemed *a priori* to be in the region of r = 0.7 to 0.8, good to very good being r = 0.8. Rohlf considers that for more than 12 OTUs (Operational Taxonomic Unit) a correlation greater than 0.5 will be statistically significant at the 99% level. The correlation is subjected to a Student's t-test to assess statistical significance. Note that the significance tests are biased because the matrices being compared are not fully independent.

Principle Components Analysis (PCA) is an ordination analysis technique that attempts to replace a large set of variables by selecting a smaller subset of variables which best summarizes the larger set. The two-fold advantage of this technique is the simplification of a large data set by highlighting significant sources of variance – in this case the characters, and the extraction of a relationship structure between the variables - in this study the species inter-relationships. The relationships between OTUs in low dimensional space (2-D or 3-D) and the efficiency of the true separation of points can be assessed by superimposing a Minimum Spanning Tree (MST) onto the first two principle component axes. Multiple methods of analysis were used because, as pointed out by Duncan & Baum (1981), different algorithms bias the results in different ways. The use of different methods of analysis allows the verification of the validity of the groups suggested by these analyses. Restricted PCA analyses on different subsets of the data allowed the selection of characters contributing the most variation separating the subgenera, sections and series. The ten most highly ranked characters (eigenvectors) for the first two components in three PCA runs (total data set, the Cicer annuals and the *Cicer* perennials) were selected and cross-referenced to the original data sets for systematic usefulness. These characters form the basis of the descriptions for the supraspecific ranks in the nomenclatural analysis.

RESULTS

Cluster analysis

The results of the cluster analysis are shown in Fig. 2. The species are divided into two clusters at the 46% level. One group, group A, contains all 9 annual species with 3 perennials: *C. canariense*, *C. incisum* and *C. atlanticum*. The second group, group B,



Fig. 2. Phenogram of all Cicer species based on 104 characters.

covers the remaining 29 species, which are all perennials. As similarity increases to 53% the taxa form 7 further clusters (numbered from 1 to 7 in Fig. 2). Three of these clusters fall under group A, and the remaining 4 into group B. The division of taxa into these clusters is as follows:

Cicer canariense and C. cuneatum.

The two varieties of *C. pinnatifidum* are most similar and these are most closely associated with *C. judaicum* and *C. bijugum*. The remaining species are *C. incisum*, *C. chorassanicum*, *C. yamashitae* and *C. atlanticum*.

Three closely related annual species: *C. arietinum*, *C. reticulatum* and *C. echino-spermum*.

Cicer floribundum, C. graecum, C. heterophyllum, C. isauricum and C. montbretii. Cicer baldshuanicum, C. mogoltavicum, C. oxyodon, C. spiroceras and C. kermanense.

Cicer paucijugum seems somewhat isolated from the other species in the cluster. *Cicer nuristanicum*, *C. songaricum*, *C. flexuosum*, *C. multijugum*, *C. microphyllum* and *C. fedtschenkoi*, and more remotely *C. korshinskyi*, *C. grande*, *C. balcaricum* and *C. anatolicum*.

Cicer acanthophyllum, C. macracanthum, C. pungens, C. tragacanthoides (including the two varieties), *C. rechingeri, C. stapfianum, C. incanum* and *C. subaphyllum*.



Fig. 3. PCA plot with MST superimposed. Clusters delineated by dotted lines and identified by I, II and III. Species names abbreviated to first four letters of the specific epithet, or in the case of subspecific epithets, the first letter of the specific name and the first three letters of the subspecific epithet, i.e. 'acan' = C. acanthophyllum; 'ttur' = C. tragacanthoides var. turcomanicum Popov.

Cophenetic correlation and principle coordinates analysis

The results of the multivariate techniques PCA and MST are shown in Fig. 3. The Mantel statistic Z gives r = -0.81, with the t-test giving t = -19.37 and p < 0.000, which indicates the result is a statistically reliable representation of the relationships between the OTUs. The PCA plot of the first two Principal Coordinates with a superimposed MST reinforces the preliminary groupings suggested by the cluster analysis. Three main clusters can be seen (identified in Fig. 3 by the roman numerals given here and delineated by a dotted line) and the membership of each is as follows:

- Cluster I: Cicer arietinum, C. atlanticum, C. bijugum, C. canariense, C. chorassanicum, C. cuneatum, C. echinospermum, C. incisum, C. judaicum, C. pinnatifidum var. anatolicum, C. pinnatifidum var. pinnatifidum, C. reticulatum and C. yamashitae.
- Cluster II: Cicer acanthophyllum, C. incanum, C. macracanthum, C. pungens, C. rechingeri, C. stapfianum, C. subaphyllum, C. tragacanthoides var. turcomanicum and C. tragacanthoides var. tragacanthoides.
- Cluster III: Cicer anatolicum, C. balcaricum, C. baldshuanicum, C. fedtschenkoi, C. flexuosum, C. floribundum, C. graecum, C. grande, C. heterophyllum, C. isauricum, C. kermanense, C. korshinskyi, C. microphyllum, C. mogoltavicum, C. montbretii, C. multijugum, C. nuristanicum, C. oxyodon, C. paucijugum, C. songaricum and C. spiroceras.

Table 3. Eigenvalues, % variance and cumulative variance a	after PCA for the first 3 components in
the analyses (PCA 1 'all taxa' - 104 characters, 43 species; P	PCA 2 'Cicer annuals' – 104 characters,
11 species; PCA 3 'Cicer perennials' - 92 characters, 32 spe	ecies).

	Component	Eigenvalue	%Variance	Cumulative Variance
	1	15.14	16.46	16.46
All axa	2	10.51	11.42	27.88
1	3	5.09	5.54	33.42
r ds	1	19.58	21.05	21.05
<i>ice</i> nua	2	16.55	17.79	38.84
an	3	14.14	15.20	54.05
r als	1	13.55	14.73	14.73
<i>ice</i>	2	9.14	9.93	24.67
C pere	3	6.79	7.39	32.05

Character selection for supraspecific descriptions

The cumulative variance accounted for in the first two component axes was relatively low in all analyses (All taxa = 28%; Annuals = 38%; Perennials = 25%). This indicates that there is a lot of 'noise' in the data set, i.e., the characters are intrinsically variable (Table 3, 4).

Some characters in PCA 1 are principally important in defining the clusters among the perennials species such as leaflet characters and corolla size. Other characters are more important in circumscribing the annuals such as rachis apex or stem height. The remaining characters such as filament length, life cycle, habit, calyx tube length, presence of a terminal leaflet and leaflet shape could be considered as potential descriptors for subgeneric groups. Within the subgenera, the characters listed for PCA 2 and PCA 3 can be considered as potential descriptors for the sections and series.

DISCUSSION

Morphological analysis and extant taxonomy

Considering the results of the cluster and PCA analyses together, one overall picture of resemblance emerges which is compared and contrasted to previous classifications. Group A in Fig. 2 and Cluster I in Fig. 3 are identical and this grouping largely corresponds to subgenus *Pseudononis* Popov. This cluster contains all the annual and 3 perennial species and is composed of 3 or 4 subclusters which could be regarded as equivalent to sectional and series divisions within the subgenus.

The first subcluster of *C. arietinum* (the crop chickpea), *C. reticulatum* (the crop progenitor) and *C. echinospermum* are closely related species (see below) and this subcluster is directly referable to series *Arietina*. This grouping is supported by nearly all previous

PCA 1 'All taxa'		PCA 2 'Cicer annuals'		PCA 3 'Cicer perennials'		
cpt1	cpt 2	cpt1	cpt2	cpt1	cpt2	
Growth cycle	Rachis apex spine	Phyllotaxy	Internodes length	Leaflet length	Leaf shape outline	
Standard length	Leaflet length	Seed length	Calyx dorsally gibbous	Rachis apex spine	Midrib tooth recurved	
Filament length	Number teeth per leaflet	Stem height	Tendril structure	Leaflet petioles	Standard length	
Leaf shape outline	Stipules laminate	Seed coat surface	Rachis apex tendril	Leaf arrangement	Standard width	
Standard width	Leaf arrangement	Leaflet base	Habit	Leaflet width	Calyx hair density	
Alae length	Rachis apex leaflet	Allozyme variation (Kazan & & Muehlbauer, 1991)	Seed coat colour	Number teeth per leaflet	Branching	
Calyx tube length	Leaflet margins serrations	Isozyme poly- morphism (Ahmad et al., 1992)	Rachis length	Stipules laminate	Form	
Habit	Leaflet petioles	Isozyme variation (Tayyar & Waines, 1996)	Seed storage proteins (Ahmad & Slinkard, 1992)	Leaflet margins serrations	Alae length	
Stem height	Leaflet width	Midrib tooth spinous	Seed proteins (Sammour, 1994)	Calyx teeth length	Leaf spacing on rachis	
Pigmentation	Leaflet shape	Calyx length	Crossability (Ladizinsky & Adler, 1976)	Calyx hair length	Peduncle lenght	

Table 4. Character list of ten characters on the first two component axes with the highest ± ranking eigenvectors for each PCA analysis.

studies and is reflected by results in crossing experiments (Ladizinsky & Adler, 1976), seed proteins (Sammour, 1994), seed storage proteins (Ahmad & Slinkard, 1992), isozyme polymorphism (Ahmad et al., 1992), isozyme polymorphism and genetic distance (Labdi et al., 1996) and karyotype analysis (Ocampo et al., 1992).

The second subcluster of *C. bijugum*, *C. judaicum* and *C. pinnatifidum* is also a well established natural grouping and biochemical and genetic research consistently confirms this association (Ladizinsky & Adler, 1976; Ahmad et al., 1992; Labdi et al., 1996; Tayyar & Waines, 1996). The taxa were divided into two series (*Macro-aristae* Maesen and *Pinnatifida* Seferova) by Seferova (1995) on the basis of the latter having more leaflets and a marginally smaller standard petal.

The third subcluster is composed of a less cohesive group of species. The close relationship between *C. chorassanicum* and *C. yamashitae* is also supported by the same biochemical and genetic data as the two clusters discussed above. Seed storage protein data (Ahmad & Slinkard, 1992; Sammour, 1994) singly place *C. chorassanicum* and *C. yamashitae* respectively within the secondary gene pool, i.e., towards series *Pinnatifida*, but isozyme polymorphism and calculated genetic distances place these two species together removed from the GP2 (Ahmad et al., 1992; Labdi et al., 1996). These two species also share the same plumule type (PII) distinct from many other species in *Cicer* (Javadi & Yamaguchi, 2004c). In contrast to the previously published supraspecific hierarchies (Van der Maesen, 1972; Seferova, 1995) that placed both taxa in subgenus *Cicer* (= *Pseudononis*) molecular phylogenies based on nrITS, trnK/matK and trnStrnG data (Van der Maesen et al., 2007) indicate that these two closely affiliated taxa should be placed in subgenus *Viciastrum*. It is also corroborated by ITS data (Frediani & Caputo, 2005). *Cicer chorassanicum* is spread over Persia and north/central Afghanistan while *C. yamashitae* is confined to Afghanistan.

An affinity between the two perennial species *C. atlanticum* and *C. incisum* is clearly indicated by this study. Although *C. atlanticum* is placed in subgenus *Viciastrum* by some molecular phylogenies (Van der Maesen et al., 2007), which tallies with Popov's allocation using his empirical-geographical system, the morphological data places *C. atlanticum* and *C. incisum* close together in subgenus *Cicer*. This is supported by ISSR (Sudupak, 2004), AFLP (Sudupak et al., 2004), allozyme (Sudupak & Kence, 2004) and RAPD (Sudupak et al., 2002) data based on *Cicer* species from Turkey. The close specific relationship between *C. atlanticum* and *C. incisum* has previously been noted by Van der Maesen (1972: 19) and Contandriopoulos et al. (1972). Although an African (Moroccan) species, *C. atlanticum* does not ally itself with the other African taxa in subgenus *Stenophylloma* (*C. canariense* and *C. cuneatum*) in the morphological or molecular analyses; it is placed much closer to *C. incisum* from the eastern Mediterranean / Persia.

Lastly, *C. canariense* and *C. cuneatum* are relatively remotely linked to each other and the other species of the cluster. Previous studies place *C. canariense* in a separate monospecific subgenus *Stenophyllum* A.G. Guerra & G.P. Lewis, on account of its geographical isolation from other *Cicer* species and distinctive characteristics. However, this study indicates a similarity to *C. cuneatum*. *Cicer cuneatum* has previously been loosely linked with the annual species, as the monospecific series *Cirrhifera* Maesen (1972) or as section *Cunecicer* Seferova (1995). The distinction of this species has also been indicated by genetic and biochemical studies (Ahmad & Slinkard, 1992; Ahmad et al., 1992; Sammour, 1994; Labdi et al., 1996; Tayyar & Waines, 1996). Seed and plumule analysis (Javadi & Yamaguchi, 2004c) also place these two species close together but at some distance from the other *Cicer* taxa. Van der Maesen (1987) described *C. canariense* as being "a vetch-like perennial" having the seedling morphology of *C. cuneatum*, this indicates close affinities with the tribe Vicieae (Nozzolillo, 1985).

Group B in Fig. 2 and Clusters II and III in Fig. 3 are the same and correspond to the species of subgenus *Viciastrum*, Cluster II, with a few exceptions, is section *Tragacanthoides* Jaub. & Spach. The nine taxa included in the latter section are all high-altitude, xerophytic species from Central Asia. The analysis indicates four subclusters within Cluster II, although the organisation of species in these groups does not precisely follow previous classifications. The grouping of *C. subaphyllum* and *C. stapfianum* is recognised as series *Subaphylla* Seferova by Seferova (1995). *Cicer incanum*, cited as a synonym of *C. macracanthum* by Seferova (1995), was placed in section *Polycicer* by Van der Maesen although he notes its affinity to *C. macracanthum* (Van der Maesen, 1972: 23). Here it is a cluster outlier, but care must be taken when interpreting its placement here as the result may be partially due to a large quantity of missing data for this species. The remaining species, *C. pungens, C. tragacanthoides* and *C. rechingeri*, are grouped together. Two of the 3 species are given monospecific status in series *Pungentia* and *Tragacanthoidea* by Seferova (1995), and she groups *C. rechingeri* with *C. macracanthum* on the basis of leaflet number, number of flowers on the inflorescences and leaflet shape. However, this distinction is not supported by the results presented here.

Groups 4, 5 and 6 (Fig. 2) and Cluster III (Fig. 3) represent the most complex and least resolved set of species in the genus *Cicer*. Many taxa have often switched taxonomic position and taxonomic rank. Popov (1929) defined series composition based on geographical distribution; but, as Coles et al. (1998) concluded, many *Cicer* species are seriously under-collected and their true pattern of distribution is incomplete. However, group B4 is geographically centred on the Greek and Turkish Aegean islands and is referable to section *Polycicer*. This grouping is supported by the cluster and ordination results as well as karyotype data; *C. graecum*, *C. heterophyllum*, *C. isauricum* and *C. montbretii* have 2n = 16, while *C. floribundum* has 2n = 14 (Contandriopoulos et al., 1972).

During the analyses certain taxa appeared to hold consistently stable key positions in the patterns of relationships (see Fig. 3) implying that they held closer affinities between subclusters than other taxa, while belonging quite clearly to independent subclusters (C. balcaricum, C. flexuosum, C. grande and C. nuristanicum). Closer study showed that these were among the species which had undergone the greatest taxonomic reclassification among previous classifications. Cicer anatolicum and C. balcaricum have been grouped together and referred to as series Anatolo-Persica (Popov) Lincz. in previous classifications, but appear quite separate in this analysis. Conversely, C. grande and C. korshinskyi are close in this analysis, which concurs with Van der Maesen's (1972) view that both are in series Flexuosa. Seferova keeps C. grande in series Flexuosa but moves C. korshinskyi to series Anatolo-Persico-Orientale. Many of the species in this group appear to form small tight groups rather than larger clusters. So, while this analysis has clustered C. kermanense, C. spiroceras and C. oxyodon with C. baldshuanicum and C. mogoltavicum, the former group of 3 species has been recognised as a distinct group (series Persica). Cicer baldshuanicum was placed with C. mogoltavicum in series Flexuosa Lincz. by Van der Maesen (1972). Seferova (1995) retained C. baldshuanicum in series Flexuosa Seferova and recombined C. mogoltavicum as C. flexuosa subsp. mogoltavicum Popov.

It is interesting to note that many species with similar geographical distribution are also linked by the results of the analyses. This either implies that the characters used here to segregate the taxa are heavily biased towards those influenced by geography or environment or they represent suites of characters which, when combined, are not independent within *Cicer. Cicer songaricum, C. microphyllum* and *C. anatolicum,* while appearing morphologically very similar, can be distinguished by their allopatric distribution: *C. songaricum* is confined to Central Asia, while *C. microphyllum* and *C. anatolicum* are from Kashmir and Turkey, respectively (Van der Maesen, 1987).

Character suites and novel taxonomic hierarchy

The multivariate analysis clearly indicated that the characters available for segregating taxa are very variable. Consequently, using only one or two features to delineate infrageneric groups will result in an indistinct, unstable hierarchy. This strongly argues for groups defined using many more characters, to account for the intrinsic variation. It also implies that morphology alone cannot be used to define the species relationships. The morphometric analyses did not reveal any novel taxonomically useful characters. Instead, the morphometric analysis allowed the definition of taxonomically useful character suites, whose variation allowed the circumscription of distinct groups. Thus, in a departure from conventional descriptions that restrict supraspecific descriptions to a few minimal characters, the subgenera, sections and series are defined by comparatively many characters.

Homoplasy (similarity not as a direct result of common ancestry) or repeated evolution of certain character traits within the genus, is clearly a disrupting factor for the development of a monophyletic infrageneric structure. Traditionally favoured characters, e.g., life cycle (annual : perennial) or rachis terminal structure (tendril : leaflet : spine) are particularly weak. By down-playing the importance of these characters in the definition of the infrageneric ranks of *Cicer*, and acknowledging the recent advances in molecular research of *Cicer*, it should be possible to construct a taxonomically useful infrageneric hierarchy that reflects interspecific relationships.

The revised classification proposed for *Cicer* is based on the morphometric analyses but take into account previously published hierarchies and the newly published molecular phylogenies.

The three subgenera *Cicer*, *Viciastrum* and *Stenophyllum* are distinguished on the basis of a suite of characters including habit (herbaceous : woody), life cycle (perennial : annual), terminal structure of rachis (tendril : leaflet : spine), leaf shape (outline parallel : outline decrescent), corolla size (standard long and broad : short and narrow) and filament length (long : short). The broad geographical distribution (African : Aegean-Mediterranean : West and Central Asia) is also important.

The sections and series in the subgenus *Cicer* are defined by the following suite of characters. Phyllotaxy (nearly opposite : alternate), stem height (< 20 cm : > 20 cm), leaf base shape (rounded cuneate to cuneate), number of leaflet pairs on rachis (3-10:1-3(-7)), midrib tooth features (± spinous : recurved : prominent), calyx teeth length (< 7 mm : > 7 mm), seed length (c. 3–4 mm : c. 6 mm), seed coat surface features (wrinkled : echinate : tuberculate). The hierarchy of the primary and secondary gene pool is acknowledged by the nomenclatural series *Cicer* and *Pinnatifida* within section *Cicer*. Seed storage proteins (Ahmad & Slinkard, 1992, Sammour, 1994) and isozyme polymorphism data (Ahmad et al., 1992; Labdi et al., 1996; Tayyar & Waines, 1996) corroborate this grouping, as do RAPD DNA analyses (Ahmad, 1999; Sudupak et al., 2002). Section *Chamaecicer* includes the two perennial species in this subgenus: *C. atlanticum* and *C. incisum*.

Subgenus Viciastrum is divided into three monophyletic sections: Annua, Polycicer and Vicioides s. ampl. Section Annua groups C. yamashitae and C. chorassanicum together. Subgenus Viciastrum section Polycicer forms a unified morphological subset within section Vicioides. The character traits defining and uniting the five taxa in Polycicer are also frequently found in various combinations in section Vicioides. However, the following character traits are unique to Polycicer: Habit (always herbaceous, flexuous

stems); branching (mostly simple), leaf outline (always equal), leaflets (petiolate, always entirely dentate); raceme (1–many-flowered), arista (terminal clavate leaflet, not spinous); distribution (below 2000 m a.s.l. in Aegean-Mediterranean). The importance of the unique eco-geographic affiliation and the molecular phylogeny (Van der Maesen et al., 2007) of these species is recognised by the rank of section.

Subgenus *Viciastrum* section *Vicioides* s. ampl. indicates a significant shift in the supraspecific taxonomic hierarchy. Both the molecular data and several morphological analyses (e.g., Van der Maesen et al., 2007) indicate that the taxa within this group show a high degree of homoplasy, with parallel developments of character traits resulting in genetically distant (Van der Maesen et al., 2007) but morphologically similar taxa, i.e., *C. oxyodon* and *C. spiroceras*. These have 'traditionally' been the taxa whose precise taxonomic position has been most disputed and unstable. To avoid the persistent non-monophyletic artificial groupings that have characterised the description of these species' relationships in the past, the section *Vicioides* has been emended and amplified, and the subsectional ranks have been abandoned.

Subgenus *Stenophyllum* is robustly supported by both the morphological and molecular studies, and includes both *C. canariense* and *C. cuneatum*. This nomenclatural relocation of *C. cuneatum* and acknowledgment of its phylogenetic origin as closer to *C. canariense* has already been suggested by several authors (viz. Frediani & Caputo, 2005; Javadi, 2004; Javadi & Yamaguchi, 2004 a, b; Van der Maesen et al., 2007).

INFRAGENERIC CLASSIFICATION OF CICER

Cicer

Cicer L. (1753). = Nochotta S.G. Gmel. = Spiroceras (Jaub. & Spach) Hutch., nom. invalid.

Subgenus Cicer

Cicer L. subg. Cicer. — Cicer L. subg. Pseudononis Popov (1929) 168, nom. illeg.; Maesen (1972) 18. — Type species: Cicer arietinum L.

Herbaceous (seldom woody at base) slender annual or perennials. Stems \pm densely pubescent with eglandular to glandular hairs. Leaf outline is equal, not decrescent. Terminal leaflet on leaves. Stipules laminate, usually smaller than leaflets. Leaflets laminate; cuneiform, obovate or elliptic; rarely flabellate, always dentate but sometimes only at apex; midrib tooth \pm prominent, \pm spinous, \pm recurved; petiolate, subsessile or sessile. Peduncles usually short (0–15 mm long) but some species with longer peduncles (> 30 mm L.) Pedicels short or long. Calyx teeth straight; calyx tube short, < 5 mm long. Standard small, 5–10(–15) mm long, and narrow, 5–10(–15) mm wide; obovate to ovate; marginate or emarginate; pubescent or rarely glabrous; blue, pink or white. Alae shortly to longly auriculate or not auriculate; alae oblong or obovate. Filaments short, < 10 mm long. Pods oblong, or ovate; pubescence glandular or eglandular; seeds angular, globular or obovate; seed coat surface tuberculate, echinate, wrinkled or smooth, \pm prominent chalazal tubercle.

Section Cicer

Cicer L. sect. Cicer. – Cicer L. sect. Arietaria Jaub. & Spach (1842) 225, p.p. nom. illeg. – Cicer L. sect. Monocicer Popov (1929) 168, p.p. nom. illeg.; Maesen (1972) 18. – Grex Imparipinnata Alef. (1859) 357, p.p. – Type species: Cicer arietinum L.

Crop gene pools: primary and secondary.

Series Cicer

Cicer L. ser. Cicer. – Cicer L. ser. Arietina Lincz. ex Seferova (1995) 101, nom. illeg. – Cicer L. ser. Arietina Lincz. (1948) 296, nom. invalidum. – Type species: Cicer arietinum L.

Annual species, stem height 20-40 cm, leaves arranged nearly opposite or alternately, 3-10 pairs of leaflets on rachis, midrib tooth spinous, prominent, recurved; leaflet bases cuneate to rounded cuneate, calyx teeth < 7 mm (rarely more), Seed length c. 6 mm, seed coat wrinkled or echinate.

Species examined: Cicer arietinum L.; C. echinospermum P.H. Davis; C. reticulatum Ladiz.

Series Pinnatifida

Cicer L. ser. *Pinnatifida* Seferova (1995) 102. — Type species: *Cicer pinnatifidum* Jaub. & Spach. *Cicer* L. ser. *Arietina* auct. non Lincz.: Maesen (1972) 18, p.p. nom. invalidum.

Annual species, stem height < 30 cm, leaves arranged nearly opposite, 2-10 pairs of leaflets on rachis, midrib tooth \pm spinous, \pm recurved, leaflet bases cuneate, calyx teeth < 7 mm (rarely more). Seed length c. 3-4 mm, seed coat echinate or tuberculate.

Note — These species represent the secondary gene pool for the crop species. This grouping is reinforced by isozyme polymorphism data (Ahmad et al., 1992; Labdi et al., 1996; Tayyar & Waines, 1996).

Species examined: Cicer bijugum Rech.f.; C. judaicum Boiss.; C. pinnatifidum Jaub. & Spach.

Section Chamaecicer

Cicer L. sect. Chamaecicer Popov (1929) 168, p.p. – Cicer L. sect. Nanopolycicer (Popov) Seferova (1995) 98. – Cicer L. subsect. Nanopolycicer Popov (1929) 169. – Cicer L. ser. Perennia Lincz. ex Maesen (1972) 19. – Cicer L. ser. Perennia Lincz. (1948) 298, nom. invalidum. – Grex Imparipinnata Alef. (1859) 357, p.p. – Type species: Cicer incisum (Willd.) K. Malý. Cicer L. ser. Arietina auct. non Lincz.: Maesen (1972) 18, p.p. nom. invalidum.

Perennial creeping species, may be slightly woody at base, stem height < 20 cm, leaves arranged opposite, 1-3(-7) pairs of leaflets on rachis, midrib tooth not spinous but can be prominent, leaflet bases cuneate, calyx teeth < 7 mm (rarely more); seed length 3-4 mm, seed coat tuberculate.

Species examined: Cicer atlanticum Coss. ex Maire; C. incisum (Willd.) K. Malý.

Subgenus Viciastrum

Cicer L. subg. *Viciastrum* Popov (1929) 168; Maesen (1972) 19. — Lectotype: *Cicer songaricum* Steph. ex DC. (designated by Seferova, 1995).

Herbaceous or woody perennial shrubs, seldom annual herbs. Terminal leaflet, spine or tendril on leaves. Stipules laminate or spiniferous, smaller or larger than leaflets. Leaf outline always decrescent. Leaflets laminate or spiniferous, cuneiform, obovate or rotund, occasionally flabellate, always dentate but sometimes only at apex, midrib tooth \pm prominent, \pm spinous, \pm recurved, petiolate, subsessile or sessile. Peduncles 16–45 mm long. Pedicels long (> 6 mm). Calyx teeth straight (rarely recurved), tube 6–8 mm long. Standard large (12–)15–17(–27) mm, and 10–16+ mm wide, obovate to ovate, marginate or emarginate, pubescent or glabrous, blue or white, seldom yellow. Alae shortly to longly auriculate, oblong, obovate or clavate. Filaments (6–)10–11+ mm long. Pods oblong, ovate or elliptic, glandular (seldom eglandular); seeds globular or obovate, tuberculate or wrinkled, \pm prominent chalazal tubercle.

Section Annua

Cicer L. sect. Annua (Maesen) Seferova (1995) 101. — Cicer L. ser. Annua Maesen (1972) 19.
 — Cicer L. sect. Chamaecicer Popov (1929) 168, p.p. excl. typus. — Cicer L. ser. Macro-aristae Maesen (1972) 18. — Grex Imparipinnata Alef. (1859) 357, p.p. — Type species: Cicer chorassanicum (Bunge) Popov.

Annual erect to creeping species, can be slightly woody at base, stem height < 20 cm, leaves arranged opposite, 1–3 pairs of leaflets on rachis, arista to 20 mm long, seed coat tuberculate.

Species examined: Cicer chorassanicum (Bunge) Popov; C. yamashitae Kitam.

Section Polycicer

Cicer L. sect. Polycicer Popov (1929) 169. — Cicer L. ser. Graeca Seferova (1995) 97. — Cicer L. ser. Anatolica Seferova (1995) 97. — Cicer L. ser. Europaeo-Anatolica Popov (1929) 169. — Grex Imparipinnata Alef. (1859) 357, p.p. — Cicer L. sect. Arietaria Jaub. & Spach (1842) 230, p.p. — Lectotype: Cicer montbretii Jaub. & Spach (designated by Seferova, 1995).

Herbaceous, erect perennials. Flexuous stems with simple to rarely secondary branching. Leaf outline equal, leaflets evenly spaced on rachis (<2 leaflets per cm), rachis laminate or tendrilous but never spiniferous, stipules laminate. Leaflets 3–10 pairs, petiolate, (6-10) > 11 by > 4 mm, margins entirely dentate, > 12 teeth per leaflet, midrib tooth \pm recurved. Raceme 1–many-flowered, arista with terminal clavate leaflet. Calyx strongly dorsally gibbous, medium to longish pubescent (hairs > 0.6 mm), calyx teeth > 9 mm long. Standard 12–15 mm long, seldom longer, 10–15 mm wide. Alae 11–15(>16) mm long. Occurs below 2000 m in the Aegean-Mediterranean.

Species examined:

Cicer floribundum Fenzl; C. graecum Orph.; C. heterophyllum Contandr., Pamukç. & Quézel; C. isauricum P.H. Davis; C. montbretii Jaub. & Spach.

Section Vicioides

Cicer L. sect. Vicioides Jaub. & Spach (1842) 230, s. ampl. emend. Davies, Maxted & Maesen.
Cicer L. sect. Spiroceras Jaub. & Spach (1842) 232. — Cicer L. sect. Tragacanthoides Jaub.
& Spach (1842) 232. — Cicer L. sect. Polycicer subsect. Macro-polycicer Popov (1929) 169.
— Cicer L. sect. Acanthocicer Popov (1929) 169. — Cicer L. ser. Anatolo-Persica (Popov) Lincz. (1948) 299. — Cicer L. ser. Anatolo-Perso-Orientalia Popov (1929) 169. — Cicer L. ser. Flexuosa

Lincz. (1948) 299, nom. invalidum. — Cicer L. ser. Flexuosa Lincz. ex Seferova (1995) 98.
— Cicer L. ser. Microphylla Lincz. (1948) 304, nom. invalidum. — Cicer L. ser. Macracantha Lincz. (1948) 307, nom. invalidum. — Cicer L. ser. Macracantha Lincz. ex Seferova (1995) 99.
— Cicer L. ser. Persica Popov (1929) 169. — Cicer L. ser. Pungentia Lincz. (1948) 306, nom. invalidum. — Cicer L. ser. Pungentia Lincz. (1948) 306, nom. invalidum. — Cicer L. ser. Pungentia Lincz. (1948) 303, nom. invalidum. — Cicer L. ser. Subaphylla Seferova (1995) 100. — Cicer L. ser. Songorica Lincz. (1948) 303, nom. invalidum. — Cicer L. ser. Subaphylla Seferova (1995) 100. — Cicer L. ser. Tragacanthoidea Lincz. (1948) 308, nom. invalidum. — Cicer L. ser. Tragacanthoidea Lincz. ex Seferova (1995) 100. — Cicer L. ser. Anatolo-Perso-Orientalia Popov subser. Anatolo-Persica Popov (1929) 170. — Cicer L. ser. Anatolo-Perso-Orientalia Popov subser. Orientalia Popov (1929) 170. — Grex Apiculata Alef. (1859) 357. — Lectotype: Cicer songaricum Steph. ex DC.: Jaub & Spach. (= C. anatolicum Alef.) (designated by Seferova, 1995).

Cicer L. ser. Flexuosa auct. non Lincz.: Maesen (1972) 19.

Cicer L. ser. Macracantha auct. non Lincz.: Maesen (1972) 20.

Cicer L. ser. Pungentia auct. non Lincz.: Maesen (1972) 20.

Cicer L. ser. Songorica auct. non Lincz.: Maesen (1972) 20.

Cicer L. ser. Tragacanthoidea auct. non Lincz.: Maesen (1972) 20.

Woody, erect to spreading perennials, straight to flexuous stems with secondary branching. Leaf outline decrescent, leaflets widely to evenly spaced along rachis (< 1 or 2 leaflets per cm), rachis laminate, tendrilous or spiniferous, stipules laminate or spiniferous. Leaflets (3-)10-15 pairs, subsessile to sessile, <5->11 by <6 mm, margins serrated to nearly entire, <5-12 teeth per leaflet, midrib tooth always recurved. Raceme 1- or 2-flowered, arista spiniferous. Calyx strongly or weakly dorsally gibbous, glabrous to shortly pubescent (hairs <0.5 mm long), calyx teeth 5-12 mm long. Standard 12-27 by 4-16 mm. Alae <10-15 mm long. Occurs between 1000-4000 m a.s.l. in West and Central Asia.

Species examined:

Cicer acanthophyllum Boriss.; C. anatolicum Alef.; C. balcaricum Galushko; C. baldshuanicum (Popov) Lincz.; C. fedtschenkoi Lincz.; C. flexuosum Lipsky; C. grande (Popov) Korotkova; C. incanum Korotkova; C. korshinskyi Lincz.; C. laetum Rassulova & Sharipova; C. luteum Rassulova & Sharipova; C. macracanthum Popov; C. microphyllum Benth.; C. multijugum Maesen; C. nuristanicum Kitam.; C. paucijugum (Popov) Nevski; C. pungens Boiss.; C. rassuloviae Lincz.; C. rechingeri Podlech; C. songaricum Steph. ex DC.; C. stapfianum Rech.f.; C. subaphyllum Boiss.; C. tragacanthoides Jaub. & Spach.

Subgenus Stenophylloma

 Cicer L. subg. Stenophylloma A.G. Guerra & G.P. Lewis (1985) 459. — Cicer L. sect. Cuneicicer Seferova (1995) 101. — Cicer L. ser. Cirrhifera Maesen (1972) 18. — Grex Cirrhifera Alef. (1859) 356. — Type species: Cicer canariense A.G. Guerra & G.P. Lewis.

Perennial or annual, slender, semi-erect to trailing or climbing herb or shrub. Terminal simple or branched tendril on upper leaves at least. Stipules laminate, smaller than leaflets. Leaf outline equal, rachis loosely rolled. Leaflets laminate, linear or obovate, margins dentate, midrib tooth prominent, spinous, \pm recurved, pubescent, subsessile. Peduncles short or absent. Pedicels short. Calyx teeth straight, tube < 5 mm. Standard short, (5–)10–12(–15) mm long and narrow, 5–10 mm wide, obovate, emarginate, pubescent, blue or pink. Alae shortly auriculate, obovate. Filaments (6–)10–11+ mm long. Pods oblong, glandular; seeds globular, tuberculate, prominent chalazal tubercle. Occurs in Africa and the Canary Islands.

Species examined: Cicer canariense A.G. Guerra & G.P. Lewis; C. cuneatum Hochst. ex A. Rich.

ACKNOWLEDGEMENTS

The authors would like to thank BM, E, K and WAG for the loan of specimens, and also Mr. and Mrs. I.J.B. Robertson and Dr. R. Davies for financial support and encouragement.

REFERENCES

- Ahmad, F. 1999. Random amplified polymorphic DNA (RAPD) analysis reveals genetic relationships among the annual Cicer species. Theor. Appl. Genet. 98: 657–663.
- Ahmad, F., P.M. Gaur & A. Slinkard. 1992. Isozyme polymorphism and phylogenetic interpretations in the genus Cicer L. Theor. Appl. Genet. 83: 620–627.
- Ahmad, F. & A. Slinkard. 1992. Genetic relationships in the genus Cicer L. as revealed by polyacrylamide gel electrophoresis of seed storage proteins. Theor. Appl. Genet. 84: 688–692.
- Alefeld, F. 1859. Ueber die Vicieen. Oesterr. Bot. Z. 9: 352-366.
- Boissier, E. 1872. Flora Orientalis. Diag. Ser. 1, 9: 130-129; 2, 2: 43-44.
- Brummitt, R.K. & C.E. Powell. 1992. Authors of Plant Names. Royal Botanic Gardens, Kew.
- Coles, S. 1993. The creation of identification aids for Cicer species. Unpublished MSc Thesis, University of Birmingham.
- Coles, S., N. Maxted & L.J.G. van der Maesen. 1998. Identification aids for Cicer (Leguminosae, Cicereae) taxa. Edinburgh J. Bot. 55: 243–265.
- Contandriopoulos, J., A. Pamukçuoğlu & P. Quézel. 1972. A propos de Cicer vivaces du pourtour Mediterranéen Oriental. Biol. Gallo-Hellen. 4-1: 3–18.
- Davis, P.H. 1970. Cicer L. In: P.H. Davis (ed.), Flora of Turkey and the Eastern Aegean Islands, Vol. 3: 267–274. Edinburgh, Edinburgh University Press.
- De Leonardis, I.S., G. Fichera & A. Zizza. 1994. Morphological study of pollens and seeds on annual taxa of the genus Cicer L. (Leguminosae). Giorn. Bot. Ital. 127: 1101–1113.
- Duncan, T. & B.R. Baum. 1981. Numerical phenetics: its uses in botanical systematics. Annual Rev. Ecol. Syst. 12: 387–404.
- Frediani, M. & P. Caputo. 2005. Phylogenetic relationships among annual and perennial species of the genus Cicer as inferred from ITS sequences of nuclear ribosomal DNA. Biol. Pl. 49, 1: 47–52.
- Hannan, R., N. Açikgöz & L.D. Robertson. 2000. Chickpeas (Cicer L.). In: N. Maxted & S.J. Bennett (eds.), Plant genetic resources of legumes in the Mediterranean: 113–122. Dordrecht, Kluwer Academic Publishing.
- Holmgren, P.K., N.H. Holmgren & L.C. Barnett (eds.). 1990. Index Herbariorium I: The herbaria of the world (ed. 8). New York: New York Botanical Garden.
- Jaubert, H.F. & E. Spach. 1842. Monografia generis Cicer. Ann. Sci. Nat., Bot. sér. 2, 18: 228-231.
- Javadi, F. 2004. Molecular phylogeny of the genus Cicer L. (Fabaceae). PhD thesis, Osaka Prefecture University, Department of Agriculture and Biological Sciences: 1–169.
- Javadi, F., M.F. Wojchiechowski & H. Yamaguchi. 2007. Geographical diversification of the genus Cicer (Leguminosae: Papilionoideae) inferred from molecular phylogenetic analyses of chloroplast and nuclear DNA sequences. Bot. J. Linn. Soc. 154: 175–186.
- Javadi, F. & H. Yamaguchi. 2004a. Interspecific relationships of the genus Cicer L. (Fabaceae) based on trnT-F sequences. Theor. Appl. Genet. 109: 317–322.
- Javadi, F. & H. Yamaguchi. 2004b. RAPD and seed coat morphology variation in annual and perennial species of the genus Cicer L. Genet. Resources Crop Evol. 51: 783–794.
- Javadi, F. & H. Yamaguchi. 2004c. A note on seed coat and plumule morphological variation in the genus Cicer L. (Fabaceae). Sci. Rep. Grad. Sch. Agric. & Biol. Sci., Osaka Pref. Univ. 56: 7–16.
- Kabir, G. & R.M. Singh. 1990. Karyotype analysis of six species of Cicer L. Bangladesh J. Bot. 19: 175–181.
- Kazan, K. & F.J. Muehlbauer. 1991. Allozyme variation and phylogenetics in annual species of Cicer L. Pl. Syst. Evol. 175: 11–21.
- Kupicha, F.K. 1977. The delimitation of the tribe Vicieae (Leguminosae) and relationships of Cicer L. Bot. J. Linn. Soc. 74: 131–162.
- Kupicha, F.K. 1981. Vicieae. In: R.M. Polhill & P.H. Raven (eds.), Advances in legume systematics: 377–381. Royal Botanic Gardens, Kew.

- Labdi, M., L.D. Robertson, K.B. Singh & A. Charrier. 1996. Genetic diversity and phylogenetic relationships among the annual Cicer species as revealed by isozyme polymorphism. Euphytica 88: 181–188.
- Ladizinsky, G. 1975. A new Cicer from Turkey. Notes Roy. Bot. Gard. Edinburgh 34: 201-202.
- Ladizinsky, G. & A. Adler. 1976. Genetic relationships among the annual species of Cicer L. Theor. Appl. Genet. 48: 197–203.
- Linczevski, I.A. 1948. Cicer L. In: V.L. Komorov (ed.), Flora of the U.S.S.R. 13: 294–309. Moskova-Leningrad, Izdatelstvo Akademii Nauk S.S.S.R.
- Linnaeus, C. 1753. Species Plantarum ed. 1: 738.
- Maxted, N. 1993. A phenetic investigation of Vicia L. subgenus Vicia (Leguminosae, Vicieae). Bot. J. Linn. Soc. 111: 155–182.
- Muehlbauer, F.J., W.J. Kaiser & C.J. Simon. 1994. Potential for wild species in cold season food legume breeding. Euphytica 73: 109–114.
- Nozzolillo, C. 1985. Seedling morphology and anatomy of 8 Cicer L. species and their taxonomic value. Canad. J. Bot. 63: 1–6.
- Ocampo, B.G., A. Venora, A. Errico, K.B. Singh & F. Saccardo. 1992. Karyotype analysis in the genus Cicer L. J. Genet. & Breed. 46: 229–240.
- Popov, M.G. 1929. The genus Cicer and its species. Trudy Prikl. Bot. 21: 1-254.
- Rajesh, P.N., V.J. Sant, V.S. Gupta, F.J. Muehlbauer & P.K. Ranjekar. 2003. Genetic relationships among annual and perennial wild species of Cicer using Inter Simple Sequential Repeat (ISSR) polymorphism. Euphytica 129: 15–23.
- Rassulova, M.R. & B.A. Sharipova (eds.). 1978. Flora Tadzhikskoi S.S.R. 5: 562-634.
- Rassulova, M.R. & B.A. Sharipova. 1992. New species of the genus Cicer from Tajikistan. Izv. Akad. Nauk Tadziksk. SSR 1: 51–52.
- Robertson, L.D., K.B. Singh, W. Erskine & A.M. Abd el Moneim. 1996. Useful genetic diversity in germplasm collections of food and forage legumes from West Asia and North Africa. Genet. Resources Crop Evol. 43: 447–460.
- Rohlf, F.J. 1995. NTSYS pc: Numerical taxonomy and multivariate analysis systems. New York, Exeter Software.
- Sammour, R.H. 1994. Seed protein homology and species relationships in the tribe Vicieae. Feddes Repert. 105: 191–196.
- Santos Guerra, A. & G.P. Lewis. 1985. A new species of Cicer (Leguminosae–Papilionoideae) from the Canary Islands. Kew Bull. 41: 459–462.
- Seferova, I.V. 1995. Konspekt sistemy roda Cicer (Fabaceae). (The synopsis of the system of genus Cicer (Fabaceae)). Bot. Zhurn. (Moscow & Leningrad) 80-8: 96–104.
- Sudupak, M. 2004. Inter and intra-species Inter Simple Sequence Repeat (ISSR) variations in the genus Cicer. Euphytica 35, 2: 229–238.
- Sudupak, M., M. Akkaya & A. Kence. 2002. Analysis of genetic relationships among perennial and annual Cicer species growing in Turkey using RAPD markers. Theor. Appl. Genet. 105, 8: 1220–1228.
- Sudupak, M., M. Akkaya & A. Kence. 2004. Genetic relationships among perennial and annual Cicer species growing in Turkey assessed by AFLP fingerprinting. Theor. Appl. Genet. 108, 5: 937–944.
- Sudupak, M. & A. Kence. 2004. Genetic relationships among perennial and annual Cicer species growing in Turkey as revealed by allozymes. Genet. Resources Crop Evol. 51, 3: 241–249.
- Tayyar, R.I. & J.G. Waines. 1996. Genetic relationships among the annual species of Cicer (Fabaceae) using isozyme variation. Theor. Appl. Genet. 92: 245–254.
- Van der Maesen, L.J.G. 1972. Cicer L., A monograph of the genus with special reference to the chickpea (Cicer arietinum L.), its ecology and cultivation. Meded. Landbouwhoogeschool Wageningen 72-10: 9–136.
- Van der Maesen, L.J.G. 1987. Origin, history and taxonomy of the chickpea. In: M.C. Saxena & K.B. Singh (eds.), The chickpea: 11–34. Wallingford, CAB International.
- Van der Maesen, L.J.G., N. Maxted, F. Javadi, S. Coles & A.M. Davies. 2007. Taxonomy of Cicer revisited. In: S.S. Yadav, R. Redden, W. Chen & B. Sharma (eds.), Chickpea breeding and management: 14–46. CABI International.

IDENTIFICATION LIST

<i>a</i> ·	
(ICPr	
Cicci.	

1 = anatolicum	12 = graecum	23 = pinnatifidum var. anatolicum
2 = arietinum	13 = incisum	24 = pinnatifidum var. pinnatifidum
3 = atlanticum	14 = isauricum	25 = pungens
4 = bijugum	15 = judaicum	26 = rechingeri
5 = canariense	16 = kermanense	27 = reticulatum
6 = chorassanicum	17 = macracanthum	28 = songaricum
7 = cuneatum	18 = microphyllum	29 = spiroceras
8 = echinospermum	19 = montbretii	30 = subaphyllum
9 = fedtschenkoi	20 = multijugum	31 = tragacanthoides var. tragacanthoides
10 = flexuosum	21 = nuristanicum	32 = tragacanthoides var. turcomanicum
11 = floribundum	22 = oxyodon	33 = yamashitae

Aitchison 740: 18 — Akhtar 706/45: 6 — Alava 7067: 23; 10591: 6 — Alston & Sandwith 1642: 19
 — Andrews R117: 5 — Archibald 2187: 1 — Aryavand, Edmondson & Miller 1420: 29 — Assadi, Edmundson & Miller 1681: 16 — Atchley 627: 12.

A. Baytop 20127a: 4 – A. Baytop & T. Baytop 2583: 19 – Biggs 13153: 16 – J. Bormüller & A. Bormüller 6634: 22; 6635: 22 – Bowes 88: 17 – Brown & Rothera 15: 18.

- Carter 542: 20 Clarke 28716B: 17 Cosson 1873 (BM-type): 3; 1876 (BM-type x2): 3.
- Daly 1246: 31 Davis 42876: 8; 43077: 24 P.H. Davis 307: 13; 4339: 15; 18119: 13 Davis & Hedge 27464: 24; 28545: 8; 29090: 23; 29179: 23; 29402: 1; 31201: 13; 31766: 1 Davis & Polunin 24751: 1 De Wilde 7041: 7.
- E00032291: 18 Edmondson 1207: 32.
- Fouroughi, Sanii & Amini 12347: 29 Furse 2568: 31; 2624: 22; 5692: 6; 8396: 6 Furse & Synge 427: 22.
- Gillet & Rawi 7669: 4.

Hedge & Wendelbo 4202: 25; 4415: 25; 4525: 25; 5225: 26; 5348: 9; 8793: 31.

Kerr 2422: 2 - Kie 2378: 25 - Kotschy 403: 30.

Lemperg 434: 12 — Litinow 388: 32 — Ludlow 356: 18 — Ludlow & Sherriff 8399: 18; 8547: 18.
Markova et al. 932: 19 — Massodegh 15: 29 — Maxted, Allkin & Khattab 4699: 19 — Maxted, Auricht & Kitiki 4843: 8; 5043: 8; 5230: 8 — Maxted, Ehrman, & Khattab 1870: 15; 1877: 15 — Maxted, Ladizinsky & Potokina 8045: 10; 8048: 10; 8062: 10 — Maxted & Sperling 8201: 28; 8234: 28 — Musselman 10329: 15.

- Noë 174: 19.
- Orphanides 495: 13.
- Pappi 138: 7; 6352: 7 Polunin et al. 695: 2; 1197: 18 Punjab Department of Agriculture Type 14: 2; Type 20: 2.
- Rabmanian 6321: 31 Rechinger 10443: 22; 18720: 25; 37351: 20; 47970: 22; 53396: 32 Roberts 566: 18 Rowe & Sproul 34: 9.
- Schimper 270 (BM-isotype): 7; 810 (BM-isotype): 7; 1545 (BM-isotype): 7 Siddiqui 4130: 18
 Southampton University 125: 18 Stainton 2672: 21; 2691: 17 Stapf 1294: 22.
- Van der Maesen 1347: 14; 2022: 33; 2023: 33; 2024: 33; 2058: 11; 2060: 11; 2100: 27; 2103: 4; 2105: 27; 2106B2: 27; 2211: 26; 2214: 26; 2765: 14; 3212: 12; Sdl. 1033-68: 10.

Wheeler Haines W1544: 4.

Zohary & Amdursky 345: 15.

From the literature: Contandriopoulos et al. (1972): C. heterophyllum; Van der Maesen 1972, 1987; Davis, 1970; Maxted, 1993; Kupicha, 1977; Coles, 1993: C. acanthophyllum; C. balcaricum; C. baldshuanicum; C. grande; C. incanum; C. korshinskyi; C. mogoltavicum; C. paucijugum; C. stapfianum.