

DISPOSITION OF RECENTLY DESCRIBED SPECIES OF *PENICILLIUM*

JENS C. FRISVAD*
ROBERT A. SAMSON** and AMELIA C. STOLK**

Hundred and twenty-two species, varieties, and new combinations of *Penicillium*, *Eupenicillium*, and *Talaromyces* described since 1977 have been studied taxonomically and screened for mycotoxin production. Only 48 taxa could be accepted: *Eupenicillium angustiporcatum*, *E. cryptum*, *E. lineolatum*, *E. limoneum*, *E. nepalense*, *E. sinaicum*, *Penicillium aethiopicum*, *P. coalescens*, *P. confertum*, *P. coprobium*, *P. coprophilum*, *P. dendriticum*, *P. eberhardtii*, *P. erythromellis*, *P. flavidostipitatum*, *P. heteromorphum*, *P. hispanicum*, *P. jugoslavicum*, *P. lapatayae*, *P. loliense*, *P. maclellaniae*, *P. macrosporum*, *P. mariaecrucis*, *P. mononematosum*, *P. nodulum*, *P. oblatum*, *P. onobense*, *P. palmae*, *P. palmense*, *P. panamense*, *P. patens*, *P. pittii*, *P. primulinum*, *P. rubefaciens*, *P. sabulosum*, *P. shennongjianum*, *P. siamense*, *P. smithii*, *P. vasconiae*, *P. von-arxii*, *P. vulpinum*, *Talaromyces assiutensis*, *T. derxii*, *T. macrosporus*, and *T. mimosinus*. Eleven varieties are recognized in *P. aurantiogriseum*, *P. chrysogenum*, *P. glandicola*, *P. griseofulvum* and *P. hirsutum*. *Paecilomyces pascuus* Pitt & Hocking and *Geosmithia viridis* Pitt & Hocking are transferred to *Penicillium*.

Since 1977 122 new names have been described in *Penicillium*, *Talaromyces* and *Eupenicillium*, which were not included in the monograph of Pitt (1980). Morphological and chemical studies of the type strains in our laboratories showed that some of the recently described species are not new to science. In this paper we report the results of our studies based on morphology and production of known mycotoxins and other secondary metabolites.

MATERIALS AND METHODS

Type and additional cultures of the recently described species of *Penicillium* and associated teleomorphs were obtained from the CAB International Mycological Institute, Kew, England and from J. A. Quintanilla, Compania de Industrias Agricolas, Valladolid, Spain, C. Ramírez, Instituto Jaime Ferrán de Microbiología, Madrid, Spain, J. I. Pitt, CSIRO Division of Food Research, Sydney, Australia, and Qi Zu-tong, Institute of Microbiology, Academia Sinica, Beijing, China.

The cultures were grown on CYA (Czapek-yeast autolysate agar), MEA (malt extract agar), YES (yeast extract-sucrose agar) (Frisvad & Filtenborg, 1983), oatmeal agar (Samson & Pitt, 1985), creatine-sucrose agar (Frisvad, 1985b) at 25°C and on CYA at 37°C (Samson & Pitt, 1985). The cultures were screened for mycotoxin production by a simple thin layer chromatography method (Frisvad & Filtenborg, 1983) and in some cases by HPLC (Frisvad, 1987).

* The Technical University of Denmark, Department of Biotechnology, Building 221, DK-2800 Lyngby, Denmark.

** Centraalbureau voor Schimmelcultures, P.O. Box 273, 3740 AG Baarn, The Netherlands.

RESULTS AND DISCUSSION

The isolates examined are listed in Table I. The status of the validly published taxa is summarized in alphabetic order in Table II. Several new names which were proposed by Pitt (1980) for known anamorphs of *Eupenicillium* and *Talaromyces*, to comply with Art. 59 of the International Botanical Code of Nomenclature are not included in this list. A list of nomina nuda in *Penicillium* and the correct identity of isolates deposited in major culture collections is given in Table III. The results of a TLC screening for mycotoxins and secondary metabolites are also listed in Tables II and III. Accepted taxa are indicated in the following text with an asterisk (*).

GENUS EUPENICILLIUM

****Eupenicillium angustiporcatum*** Takada & Udagawa in Trans. mycol. Soc. Japan 24: 143. 1983.

The ex-type culture produces only a few reduced conidiophores and, in spite of many attempts, no teleomorph could be observed in culture. According to Takada & Udagawa's description, the ascospores are ornamented with two prominent well-separated equatorial ridges and the valves show several low ribs. The species is probably related to *E. lineolatum*.

****Eupenicillium cryptum*** Gochenaur in Mycotaxon 26: 349. 1986.

According to the description, *E. cryptum* is probably close to *E. javanicum*. It differs morphologically in producing ascospores with prominent equatorial ridges. The type strain shows an unstable colonial morphology and a restricted carbon nutrition. It does not produce the teleomorph any more.

****Eupenicillium limoneum*** Gochenaur & Zlattner apud Stolk & Samson in Stud. Mycol. 23: 100. 1983.

The description of this species was included by Stolk & Samson (1983).

****Eupenicillium lineolatum*** Udagawa & Horie in Mycotaxon 5: 493. 1977.

Eupenicillium lineolatum was reduced to a variety of *E. javanicum* by Stolk & Samson (1983), but we now regard the taxon as a distinct species (Frisvad & al., 1990a).

****Eupenicillium nepalense*** Takada & Udagawa in Trans. mycol. Soc. Japan 24: 146. 1983.

Eupenicillium nepalense is very close to *E. inusitanum* Scott but the penicilli of the latter species are biverticillate rather than monoverticillate and the ascospores of *E. nepalense* are smaller than those of *E. inusitanum* and *E. fractum*. The three taxa may be conspecific, and a detailed chemical study has still to be done.

****Eupenicillium sinaicum*** Udagawa & Ueda in Mycotaxon 14: 266. 1982.

This species was discussed and accepted by Stolk & Samson (1983).

GENUS *PENICILLIUM**Penicillium* subgenus *Aspergilloides*

Penicillium alicantinum Ramírez & Martínez in Mycopathologia 72: 185. 1980.

Penicillium alicantinum is a synonym of *P. citreonigrum*, and this is further confirmed by the production of citreoviridin by all isolates of both taxa. Sclerotia were observed in the ex-type culture of *P. alicantinum* but also observed by Wicklow (1984) in an atypically vesiculate strain of *P. citreonigrum* producing citreoviridin.

Penicillium brevissimum Rai & Wadhvani in Curr. Sci. 45: 192. 1976.

The ex-type culture of *P. brevissimum* only produces few cylindrical conidia, but fits *P. capsulatum* Raper & Fennell and is therefore considered a synonym of this taxon.

Penicillium gallaicum Ramírez & al. in Mycopathologia 72: 30. 1980.

Like *P. alicantinum*, *P. gallaicum* is considered to be a synonym of *P. citreonigrum*. The type culture of *P. gallaicum* produces citreoviridin and forms sclerotia.

Penicillium gerundense Ramírez & Martínez in Mycopathologia 72: 182. 1980.

Because of its broadly ellipsoidal smooth-walled conidia, we consider *P. gerundense* a synonym of *P. dierckxii* Biourge.

Penicillium grancanariae Ramírez & al. in Mycopathologia 66: 79. 1978.

Penicillium grancanariae is conspecific with *P. thomii* Maire, differing slightly by the formation of rough-walled conidia with transverse striations.

**Penicillium heteromorphum* Kong & Qi in Mycosystema 1: 107. 1988.

Penicillium heteromorphum strongly resembles species such as *P. cinereoatrum* Chalabuda, *P. arabicum* Baghdadi, *P. griseolum* G. Smith, and *P. dimorphosporum* Swart, characterized by the initial production of grey, smooth, subglobose to ellipsoidal conidia that later become globose and rough. According to the original description, *P. heteromorphum* differs from these species by its inability to utilize nitrate and no growth at 37°C. However, the cultures ex type kept at CBS and CMI all grew at 25°C and 37°C and reached a diameter of 10–12 mm on CYA after one week. *Penicillium heteromorphum* shares the inability to utilize nitrate with *P. griseolum*. A more detailed study is needed to elucidate its taxonomic position.

**Penicillium hispanicum* Ramírez & al. in Mycopathologia 66: 77. 1978.

Frisvad & al. (1990c) have pointed out that *P. hispanicum* is the first available name for Raper & Thom's (1949) concept of *P. implicatum*, because the type of the latter species belongs in *P. citrinum* Thom. *Penicillium hispanicum* produces many specific secondary metabolites distinguishing it from other monoverticillate species.

**Penicillium jugoslaviicum* Ramírez & Muntanola-Cvetkovic in Mycopathologia 88: 65. 1984.

The species is reminiscent of *P. bilaiae* and *P. charlesii*, but differs by its faster growth at Czapek agar.

Table I. Taxa of *Penicillium*, *Eupenicillium*, and *Talaromyces*, described since 1977, and isolates examined.

Name	Culture
<i>P. aethiopicum</i>	IMI 285624 (T)
<i>P. alicantinum</i>	IMI 253789 = CBS 164.81 (T) ¹
<i>P. allii</i>	CBS 131.89 (T), CBS 188.88
<i>E. angustiporcatum</i>	CBS 202.84 (T)
<i>P. aragonense</i>	IMI 253790 = CBS 171.81 (T)
<i>T. assiutensis</i>	CBS 147.78 (T)
<i>P. asturianum</i>	IMI 253788 = CBS 173.81 (T)
<i>P. aurantioflammiiferum</i>	CBS 165.81 (T)
<i>P. aurantiogriseum</i> var. <i>melanoconidium</i>	IMI 321503 (T)
<i>P. aurantiogriseum</i> var. <i>neoechinulatum</i>	IMI 296937 = NRRL 13486 (T)
<i>P. aurantiogriseum</i> var. <i>polonicum</i>	CBS 222.28 (T)
<i>P. brevicompactum</i> var. <i>magnum</i>	IJFM 5954 (T)
<i>P. brevissimum</i>	CBS 763.68 (T)
<i>P. brunneostoloniferum</i>	CBS 317.50 (T)
<i>P. burgense</i>	CBS 325.89 (T)
<i>P. caeruleus</i>	Q 1147 (T), Q 1152, Q 1155, Q 1161, Q 1300 ²
<i>P. castellae</i>	CBS 272.83 = Q 1012 (T), Q 1024, Q 1036, Q 1349
<i>P. castellanense</i>	IMI 253791 = CBS 170.81 (T)
<i>P. chalybeum</i>	FRR 2660 = CBS 254.87 (T), FRR 2659, FRR 2658
<i>P. chrysogenum</i> var. <i>dipodomyis</i>	IMI 296926 = NRRL 13485 (T)
<i>P. cieglerei</i>	IJFM 7673 = CBS 275.73 (T)
<i>P. cluniae</i>	CBS 326.89 (T)
<i>E. cryptum</i>	ATCC 60138 = CBS 271.89 (T)
<i>P. coalescens</i>	CBS 104.83 = Q 1138 (T)
<i>P. confertum</i>	CBS 171.87 = IMI 296930 (T)
<i>P. coprobium</i>	IMI 293209 (T)
<i>P. coprophilum</i>	CBS 477.75
<i>P. cordubense</i>	CBS 162.81 (T)
<i>P. corynephorum</i>	FRR 2663 = CBS 256.87 (T), FRR 2676
<i>P. dendriticum</i>	CBS 660.80 = FRR 1885 (T), CBS 191.89
<i>T. derxii</i>	NHL 2982 = CBS 413.89 (T) + NHL 2981 = CBS 412.89 (T)
<i>P. eberhardtii</i>	- -
<i>P. erythromellis</i>	CBS 644.80 (T)
<i>P. fagi</i>	CBS 689.77 (T)
<i>P. flavidostipitatum</i>	CBS 202.87 = IJFM 7824 (T)
<i>P. gaditanum</i>	IMI 253792 = CBS 169.81 (T)
<i>P. gallaicum</i>	IMI 253794 = CBS 333.79 (T)
<i>P. gerundense</i>	IMI 253804 = CBS 334.79 (T)
<i>P. glandicola</i> var. <i>confertum</i>	IMI 296930 = NRRL 13488 (T)

¹ T: culture ex type.² Q: from the collection of J.A. Quintanilla.

Name	Culture
<i>P. glandicola</i> var. <i>glandicola</i>	CBS 333.48
<i>P. glandicola</i> var. <i>glaucovenetum</i>	IMI 293197 (T)
<i>P. glandicola</i> var. <i>mononematosum</i>	IMI 296925 = NRRL 13482 (T)
<i>T. gossypii</i>	CBS 645.80 (T)
<i>P. granatense</i>	IMI 253795 = CBS 166.81 (T)
<i>P. grancanariae</i>	IMI 253783 = CBS 687.77 (T)
<i>P. granulatum</i> var. <i>globosum</i>	IMI 299049 (T), IMI 297543
<i>P. griseofulvum</i> var. <i>dipodomyicola</i>	IMI 296935 = NRRL 13487 (T)
<i>P. heteromorphum</i>	AS 3.4525 = CBS 226.89 (T)
<i>P. hirsutum</i> var. <i>albocoremium</i>	IMI 285511
<i>P. hirsutum</i> var. <i>allii</i>	CBS 131.89 (T)
<i>P. hirsutum</i> var. <i>hordei</i>	CBS 701.68 (T)
<i>P. hirsutum</i> var. <i>venetum</i>	IMI 321520
<i>P. hispalense</i>	- -
<i>P. hispanicum</i>	CBS 691.77 (T)
<i>P. ilerdanum</i>	IMI 253793 = CBS 335.79 (T)
<i>P. jugoslavicum</i>	IJFM 7785 = CBS 192.87 (T)
<i>P. lacus-sarmientei</i>	IJFM 19078 = CBS 685.85 (T)
<i>P. lapatayae</i>	IJFM 19012 = CBS 203.87 (T)
<i>E. limoneum</i>	CBS 650.82 (T)
<i>E. lineolatum</i>	CBS 188.77 (T)
<i>P. loliense</i>	CBS 643.80 (T)
<i>P. maclellanniae</i>	IJFM 7852 = CBS 196.81 (T), CBS 197.81, CBS 198.81
<i>T. macrosporus</i>	CBS 317.63
<i>P. malacaense</i>	IMI 253801 = CBS 160.81 (T)
<i>P. mali</i>	CBS 500.73
<i>P. mariaecrucis</i>	CBS 270.83 (T), Q 1022, Q 1049, Q 1118
<i>P. mediolanense</i>	ATCC 44200 = IJFM 7812 (T)
<i>P. melanochlorum</i>	CBS 487.75 (T), CBS 140.86, CBS 141.86 - 146.86
<i>P. michaelis</i>	CBS 144.83, Q 1150
<i>T. mimosinus</i>	CBS 659.80 (T)
<i>P. mononematosum</i>	CBS 172.87 = IMI 296925 (T)
<i>P. murcianum</i>	CBS 161.81 = IMI 253800
<i>E. nepalense</i>	CBS 203.84
<i>P. nodulum</i>	AS 3.4524 = CBS 227.89
<i>P. nordicum</i>	IJFM 7813 (T)
<i>P. oblatum</i>	FRR 2234 = CBS 258.87 (T), FRR 2233
<i>P. olivicolor</i>	CBS 246.32 (T)
<i>P. onobense</i>	IMI 253787 = CBS 174.81 (T)
<i>P. ovetense</i>	CBS 163.81 (T)
<i>P. palmae</i>	CBS 442.88 (T), CBS 829.88
<i>P. palmense</i>	CBS 336.79 (T)
<i>P. panamense</i>	CBS 128.89 = IMI 297546 (T), IMI 297558 = CBS 129.89
<i>P. patens</i>	FRR 2661 = CBS 260.87 (T)

Name	Culture
<i>P. pittii</i>	CBS 139.84 = Q 1240 (T)
<i>P. primulinum</i>	CBS 321.48 = NRRL 1074 (T)
<i>P. pulvillorum</i> var. <i>echinulatum</i>	- -
<i>P. rademiricii</i>	CBS 140.84 = Q 1248 (T)
<i>P. radiatolobatum</i>	CBS 340.79 = IJFM 7845 (T)
<i>P. resinae</i>	CBS 324.83 (T)
<i>P. roqueforti</i> var. <i>carneum</i>	IMI 293204 (T)
<i>P. rubefaciens</i>	CBS 145.83 = Q 1133 (T)
<i>P. sabulosum</i>	FRR 2743 = CBS 261.87 (T)
<i>P. sajarovii</i>	CBS 277.83 = IJFM 7674 = Q 1099 (T)
<i>P. samsonii</i>	CBS 137.84 = Q 1032 (T), Q 1100
<i>P. severskii</i>	IJFM 19000 = CBS 438.88 (T)
<i>P. shennongjianum</i>	AS 3.4526 = CBS 228.89 (T)
<i>P. siamense</i>	CBS 475.88 (T)
<i>E. sinaicum</i>	CBS 279.82 (T)
<i>P. solitum</i> var. <i>crustosum</i>	IMI 91917 (T)
<i>P. terraconense</i>	IMI 283803 = CBS 177.81 (T)
<i>P. turolense</i>	CBS 176.81 (T)
<i>P. turrispainenae</i>	CBS 204.87 (T), CBS 686.85
<i>P. vaccaeorum</i>	IJFM 7756 = Q 1134 = CBS 148.83 (T)
<i>P. valentinum</i>	IMI 253789 = CBS 338.79 (T)
<i>P. vasconiae</i>	IMI 253786 = CBS 339.79 (T)
<i>P. vonarxii</i>	CBS 348.51 (T)
<i>P. vulpinum</i>	CBS 126.63
<i>P. zacinthae</i>	CBS 178.81 (T)
<i>Geosmithia viridis</i>	FRR 1963 = CBS 252.87 (T)
<i>Paecilomyces pascuus</i>	FRR 1925 = CBS 253.87 (T)

Penicillium lacus-sarmientei Ramírez in Mycopathologia 96: 29. 1986.

Penicillium lacus-sarmientei is regarded as a faster growing variant of *P. roseopurpureum* Dierckx. The ex-type culture produced beta-hydroxycurvularin and roseopurpurin. The production of this compound has been reported by Turner & Aldridge (1983) for *P. roseopurpureum*, and roseopurpurin for *P. roseopurpureum* (Posternak, 1940) and from *P. carminoviolaceum* (Hind, 1940 a and b).

**Penicillium lapatayae* Ramírez in Mycopathologia 91: 97. 1985.

Penicillium lapatayae is a distinctive sclerotial species and it is accepted as new. It has a certain resemblance to the anamorph of *E. pinetorum*.

Penicillium malacaense Ramírez & Martínez in Mycopathologia 72: 186. 1980.

Examination of the ex-type culture showed that *P. malacaense* is a synonym of the variable *P. restrictum* Gilman & Abbott.

****Penicillium nodulum*** Kong & Qi in *Mycosystema* 1: 108. 1988.

Penicillium nodulum is characterized by its ellipsoidal smooth conidia, its dark green reverse on all substrates, its good growth on creatine-sucrose agar and restricted growth on all media (7–17 mm diam. after one week at 25°C). It is therefore accepted as a good species.

Penicillium ovetense Ramírez & Martínez in *Mycopathologia* 74: 39. 1981.

Penicillium ovetense is regarded as a synonym of *P. phoeniceum*. The illustration of this species is similar to that of a strain (CBS 583.68) of *E. cinnamopurpureum* Scott & Stolk (compare Ramírez, 1982: 318 and Stolk & Samson, 1983: 63, fig. 30 e).

****Penicillium palmense*** Ramírez & al. in *Mycopathologia* 66: 80. 1978 (as '*P. palmensis*').

This species resembles *P. thomii* by its ellipsoidal conidia, but it also shares characters with isolates of the variable *P. glabrum*. For the time being, we accept this taxon, but a more detailed study is necessary to determine its identity.

****Penicillium patens*** Pitt & Hocking in *Mycotaxon* 22: 205. 1985.

Penicillium patens resembles *P. thomii*. The conidia of *P. patens* are smooth (like in *P. quercetorum* Baghdadi, another synonym of *P. thomii*), but colony colours are quite different from those of typical *P. thomii*. A more detailed biochemical study of these species should be performed before a final conclusion is drawn.

Penicillium terraconense Ramírez & Martínez in *Mycopathologia* 72: 187. 1980.

The drawings and description of *P. terraconense* indicate that it is a monoverticillate species with small conidia and rough stipes. However, the type culture of this species was contaminated, and is a typical *P. digitatum*. Since no holotype material was designated, this taxon is considered to be of doubtful identity and the name can be discarded as being invalidly described.

Penicillium vaccaeorum Quintanilla in *Mycopathologia* 80: 77. 1982.

Like *P. lacus-sarmientei*, *P. vaccaeorum* is considered to be a fast-growing isolate of *P. roseopurpureum*.

Penicillium valentinum Ramírez & Martínez in *Mycopathologia* 72: 183. 1980.

Penicillium valentinum strongly resembles non-sclerotial isolates (e.g. CBS 338.61) of *P. thomii* and it is therefore regarded as a synonym of this species.

Penicillium subgenus Furcatum

Penicillium aragonense Ramírez & Martínez in *Mycopathologia* 74: 41. 1981.

The drawing of this taxon suggests a typical *P. oxalicum*. However, cultures derived from the type and received from CMI were *P. glabrum* and may have been confused. Since no holotype material was designated, this taxon is considered to be of doubtful identity and the name can be discarded as being invalidly described.

Penicillium asturianum Ramírez & Martínez in Mycopathologia 74: 42. 1981.

This species is regarded as a synonym of *P. oxalicum*. The ex-type culture produces the typical profile of secondary metabolites from *P. oxalicum* including secalononic acid D, oxaline and bluish fluorescent compounds.

Penicillium burgense Quintanilla in Mycopathologia (in press).

The profile of secondary metabolites and morphology of the ex-type culture is identical with *Eupenicillium lapidosum*.

Penicillium caerulescens Quintanilla in Mycopathologia 82: 101. 1983.

Penicillium caerulescens is a synonym of *P. raciborskii* sensu stricto. Isolates of *P. raciborskii*, including the ex-type culture of *P. caerulescens*, are very good producers of mycophenolic acid.

Penicillium castellae Quintanilla in Avian. Nutr. Mej. anim. Alim. 23: 336. 1982.

This is a typical *P. raistrickii*. Quintanilla (l.c.) stated that the isolates produce griseofulvin and penicillic acid and this was confirmed in this study. These two mycotoxins are also produced by the ex-type and all other isolates investigated of *P. raistrickii*.

Penicillium castellanense Ramírez & Martínez in Mycopathologia 74: 46. 1981.

Penicillium castellanense is morphologically and biochemically identical with *P. matriti*. The ex-type culture is a good producer of penicillin and penicillic acid (Frisvad & Emborg, unpubl.).

Penicillium chalybeum Pitt & Hocking in Mycotaxon 22: 204. 1985.

The ex-type culture of *P. chalybeum* produces sclerotia on MEA, a feature not recorded by Pitt & Hocking (l.c.). It is regarded here as a synonym of *Eupenicillium terrenum*.

Penicillium cieglei Quintanilla in Avian. Nutr. Mej. anim. Alim. 23: 338. 1982 (as '*P. cieglei*').

Penicillium cieglei is a typical *P. pulvillorum*. Some isolates of *P. pulvillorum*, such as the ex-type culture of *P. cieglei* and isolates assigned to *P. novae-caledoniae*, have a bright red reverse colour and often a slow growth rate on MEA.

Penicillium cluniae Quintanilla in Mycopathologia (in press).

Penicillium cluniae is a synonym of *P. cremeogriseum* Chalabuda: among other features, it shares a fast growth rate at 37°C and the production of brefeldin A.

Penicillium corynephorum Pitt & Hocking in Mycotaxon 22: 202. 1985.

Penicillium corynephorum is considered conspecific with *P. smithii* Quintanilla though it differs from it by less roughened conidiophore stipes. The type cultures of both species produce the same profile of secondary metabolites (including citreoviridin) and they have identical growth rates and conidial colours.

Penicillium fagi Martínez & Ramírez in Mycopathologia 63: 57. 1978.

Penicillium fagi, a good producer of mycophenolic acid, is the same as *P. raciborskii* sensu stricto. Like *P. caeruleum*, *P. fagi* produces a glaucous-black pigment in the reverse on MEA after 1–3 weeks storage at 0°C.

**Penicillium flavidostipitatum* Ramírez & González in Mycopathologia 88: 3. 1984.

Penicillium flavidostipitatum morphologically resembles *P. brasilianum* Batista apud Batista & Maia. However, the two species markedly differ in their profiles of secondary metabolites, growth rates and conidial colour.

Penicillium granatense Ramírez & al. in Mycopathologia 72: 31. 1980.

The morphology, profile of secondary metabolites and colony characteristics of *P. granatense* are similar to *P. janczewskii*, and therefore the species is considered a further synonym of that species (also compare Fassatiová & Kubatová, 1990).

**Penicillium macleanianae* Yip in Trans. Br. mycol. Soc. 77: 202. 1981.

In agreement with Ramírez (1985), we consider this species to be a distinct taxon characterized by the fast growth and conspicuously ornamented conidia.

**Penicillium mariaecrucis* Quintanilla in Avian. Nutr. Mej. anim. Alim. 23: 334. 1982.

Quintanilla (l.c.) described this sclerotial species as having strongly inflated metulae and phialides, but in our cultures of *P. mariaecrucis* the conidiophores, phialides, and conidia duplicate those of *P. pulvillorum*, though some atypical inflated structures were also observed. *Penicillium mariaecrucis* was found to be a good producer of xanthomegnin and viomelisin. These nephrotoxins are also produced by *P. simplicissimum*, a species closely related to *P. pulvillorum*. *Penicillium pulvillorum* itself produces penicillic acid and pulvilloric acid, while *P. mariaecrucis* produces the naphthoquinones mentioned above, so there is a marked chemical difference between the two taxa. Also the dark reddish brown colonies of *P. mariaecrucis* are different from the quite weakly coloured strains of *P. pulvillorum*. The status of this species is therefore difficult.

Penicillium michaelis Quintanilla in Mycopathologia 80: 79. 1982.

Penicillium michaelis is in all aspects a typical *P. soppii*.

Penicillium murcianum Ramírez & Martínez in Mycopathologia 74: 37. 1981.

Penicillium murcianum is identical with *P. canescens* Sopp. Isolates like *P. murcianum* resemble those intermediate between *P. canescens* and *P. janczewskii* Zaleski described by Pitt (1980).

Penicillium jensenii (finely roughened conidia, smooth-walled stipes) through *P. canescens* (smooth to finely roughened conidia, rough stipes) to *P. janczewskii* (very rough conidia, smooth to finely roughened stipes) form a continuum of species, nearly all producing the same total profile of metabolites.

Table II. The status of taxa of *Penicillium* and their teleomorphs, described since 1977, and their production of known mycotoxins.

New taxa	Synonym of	Mycotoxins produced
<i>P. aethiopicum</i>	- -	Viridicatumtoxin
<i>P. alicantinum</i>	<i>P. citreonigrum</i>	Citreoviridin
<i>P. allii</i>	<i>P. hirsutum</i> var. <i>allii</i>	Roquefortine C, meleagrins
<i>E. angustiporcatum</i>	- -	-
<i>P. aragonense</i>	<i>P. oxalicum</i>	NT ³
<i>T. assiutensis</i>		Glauconic acid
<i>P. asturianum</i>	<i>P. oxalicum</i>	Secalonic acid D, oxaline
<i>P. aurantioflammiferum</i>	<i>P. islandicum</i>	Emodin, skyrin, luteoskyrin, rugulosin
<i>P. aurantiogriseum</i> var. <i>melanoconidium</i>	- -	Penicillic acid, oxaline, penitrem A, verrucosidin
<i>P. aurantiogriseum</i> var. <i>neoechinulatum</i>	- -	Cyclopenin, viridicatin, penicillic acid
<i>P. aurantiogriseum</i> var. <i>polonicum</i>	- -	Penicillic acid, verrucosidin
<i>P. brevicompactum</i> var. <i>magnum</i>	<i>P. olsonii</i>	- ⁴
<i>P. brevissimum</i>	<i>P. capsulatum</i>	-
<i>P. brunneostoloniferum</i>	<i>P. brevicompactum</i>	Raistrick phenols, mycophenolic acid, brevianamide A and B
<i>P. burgense</i>	<i>E. lapidosum</i>	-
<i>P. caerulea</i>	<i>P. raciborskii</i>	Mycophenolic acid
<i>P. castellae</i>	<i>P. raistrickii</i>	Penicillic acid, griseofulvin
<i>P. castellonense</i>	<i>P. matriri</i>	Penicillin, penicillic acid
<i>P. chalybeum</i>	<i>E. terreum</i>	-
<i>P. chrysogenum</i> var. <i>dipodomys</i>	- -	Penicillin
<i>P. cieglerei</i>	<i>P. pulvillorum</i>	Penicillic acid
<i>P. cluniae</i>	<i>P. cremeogriseum</i>	Brefeldin A
<i>E. cryptum</i>	- -	-
<i>P. coalescens</i>	- -	-
<i>P. confertum</i>	- -	Meleagrins
<i>P. coprobium</i>	- -	Patulin
<i>P. coprophilum</i>	- -	Griseofulvin, roquefortine C, meleagrins
<i>P. cordubense</i>	<i>P. aurantiogriseum</i>	Penicillic acid, xanthomegnin, viomellein, viridicatin
<i>P. corynephorum</i>	<i>P. smithii</i>	Citreoviridin
<i>P. damascenum</i>	<i>P. westlingii</i>	Citrinin
<i>P. dendriticum</i>	- -	Mitorubrinic acid, secalonic acid D
<i>T. derxii</i>	- -	-
<i>P. eberhardtii</i>	- -	NT
<i>P. erythromellis</i>	- -	-

³ NT: not tested.⁴ -: no known mycotoxins produced.

New taxa	Synonym of	Mycotoxins produced
<i>P. fagi</i>	<i>P. raciborskii</i>	Mycophenolic acid
<i>P. flavidostipitatum</i>	- -	
<i>P. gaditanum</i>	<i>P. minioluteum</i>	Mitorubrinic acid, mitorubrin, mitorubrinol
<i>P. gallaicum</i>	<i>P. citreonigrum</i>	Citreoviridin
<i>P. gerundense</i>	<i>P. restrictum</i>	-
<i>P. glandicola</i> var. <i>confertum</i>	<i>P. confertum</i>	Meleagrins
<i>P. glandicola</i> var. <i>glandicola</i>	- -	Penitrem A, patulin, roquefortine C
<i>P. glandicola</i> var. <i>glaucovenetum</i>		
<i>P. glandicola</i> var. <i>mononematosum</i>	<i>P. mononematosum</i>	Viriditoxin, cyclopaldic acid, isochromantoxin, verrucologen
<i>T. gossypii</i>	<i>T. assiutensis</i>	Glauconic acid
<i>P. granatense</i>	<i>P. janczewskii</i>	Griseofulvin, penicillic acid, penitrem A
<i>P. grancanariae</i>	<i>P. thomii</i>	-
<i>P. granulatum</i> var. <i>globosum</i>	<i>P. glandicola</i>	NT
<i>P. griseofulvum</i> var. <i>dipodomyicola</i>	- -	Griseofulvin, patulin, cyclopiazonic acid
<i>P. heteromorphum</i>	- -	NT
<i>P. hirsutum</i> var. <i>albocoremium</i>	- -	Citrinin, roquefortine C, terrestric acid, meleagrins
<i>P. hirsutum</i> var. <i>allii</i>	- -	Roquefortine C, meleagrins
<i>P. hirsutum</i> var. <i>hordei</i>	- -	Roquefortine C, terrestric acid
<i>P. hirsutum</i> var. <i>venetum</i>	- -	Roquefortine C, terrestric acid
<i>P. hispalense</i>	<i>P. hirsutum</i> ?	NT
<i>P. hispanicum</i>	- -	-
<i>P. ilerdanum</i>	<i>P. piceum</i>	Rugulosin
<i>P. jugoslavicum</i>	- -	-
<i>P. lacus-sarmientei</i>	<i>P. roseopurpureum</i>	Beta-hydroxycurvarin, roseo purpurin
<i>P. lapatayae</i>	- -	-
<i>E. limoneum</i>	- -	-
<i>E. lineolatum</i>	- -	-
<i>P. loliense</i>	- -	-
<i>P. maclennaniae</i>	- -	-
<i>T. macrosporus</i>	- -	Duclauxin
<i>P. malacaense</i>	<i>P. restrictum</i>	-
<i>P. mali</i>	<i>P. solitum</i>	Cyclophenin
<i>P. mariaecrucis</i>	- -	Xanthomegnin, viomellein
<i>P. mediolanense</i>	<i>P. verrucosum</i>	Ochratoxin A
<i>P. melanochlorum</i>	<i>P. solitum</i>	Cyclophenin
<i>P. michaelis</i>	<i>P. soppii</i>	Terrein, asperentin
<i>T. mimosinus</i>	- -	-

New taxa	Synonym of	Mycotoxins produced
<i>P. mononematosum</i>	- -	-
<i>P. murcianum</i>	<i>P. canescens</i>	Griseofulvin, penicillic acid
<i>E. nepalense</i>	- -	-
<i>P. nodulum</i>	- -	-
<i>P. nordicum</i>	<i>P. verrucosum</i>	Ochratoxin A and B
<i>P. oblatum</i>	- -	-
<i>P. olivicolor</i>	<i>P. aurantiogriseum</i> var. <i>viridicatum</i>	Brevianamide A, viridicatin
<i>P. onobense</i>	- -	Brefeldin A
<i>P. ovetense</i>	<i>P. phoeniceum</i>	-
<i>P. palmae</i>	- -	Mitorubrin, mitorubrinol, mitorubrinol acetat
<i>P. palmense</i>	?	-
<i>P. panamense</i>	- -	Mitorubrin, mitorubrinic acid, vermicellin
<i>P. patens</i>	- -	-
<i>P. pittii</i>	- -	-
<i>P. primulinum</i>	- -	-
<i>P. pulvillorum</i> var. <i>echinulatum</i>	<i>E. zonatum</i>	NT
<i>P. rademiricii</i>	<i>P. diversum</i>	-
<i>P. radiatolobatum</i>	<i>P. canescens</i>	-
<i>P. resinae</i>	<i>P. asperosporum</i>	-
<i>P. roqueforti</i> var. <i>carneum</i>	- -	Patulin, roquefortine C, mycophenolic acid
<i>P. rubefaciens</i>	- -	-
<i>P. sabulosum</i>	- -	-
<i>P. sajarovii</i>	? <i>P. cremeogriseum</i>	Brefeldin A
<i>P. samsonii</i>	<i>P. minioluteum</i>	Mitorubrins
<i>P. severskii</i>	<i>P. soppii</i>	Terrein
<i>P. shennongjianum</i>	- -	-
<i>P. siamense</i>	- -	Mitorubrins
<i>E. sinaicum</i>	- -	-
<i>P. smithii</i>	- -	Citreoviridin
<i>P. solitum</i> var. <i>crustosum</i>	<i>P. crustosum</i>	NT
<i>P. terraconense</i>	<i>P. digitatum</i>	-
<i>P. turolense</i>	<i>P. westlingii</i>	Citrinin
<i>P. turrispainenae</i>	<i>P. namyslowskii</i>	-
<i>P. vaccaeorum</i>	<i>P. roseopurpureum</i>	Roseopurpurin, beta-hydroxy-curvularin
<i>P. valentinum</i>	<i>P. thomii</i>	-
<i>P. vasconiae</i>	- -	-
<i>P. vonarxii</i>	- -	-
<i>P. vulpinum</i>	- -	Patulin, roquefortine C
<i>P. zacinthae</i>	<i>P. allahabadense</i>	Rugulosin
<i>Geosmithia viridis</i>	<i>P. viride</i>	-
<i>Paecilomyces pascuus</i>	<i>P. pascuum</i>	-

Penicillium novae-caledoniae G. Smith var. *album* Ramírez & Martínez in Mycopathologia 74: 47. 1981.

Penicillium novae-caledoniae G. Smith (the type culture was lost but another representative isolate is IMI 140441) and its variety *album* (IJFM 7181) are both considered as synonyms of *P. pulvillorum*.

**Penicillium onobense* Ramírez & Martínez in Mycopathologia 74: 44. 1981.

The ex-type culture of *P. onobense* morphologically resembles *P. brasilianum*, but the taxon has different secondary metabolites. Being a good producer of brefeldin A, *P. onobense* resembles *E. ehrlichii* (= *E. brefeldianum*), but the anamorph of the latter has subglobose conidia without striations and less roughened conidiophore stipes.

Penicillium pulvillorum Turfitt var. *echinulatum* Basu & Mehrotra in Nova Hedwigia 27: 786. 1976.

The ex-type culture (CBS 654.82) of this variety was lost and a dried specimen was never prepared. Stolk & Samson (1983) considered this variety as a synonym of the anamorph of *E. javanicum* (van Beyma) Stolk & Scott var. *javanicum*, under which they also synonymized *E. zonatum* Hodges & Perry. Recently Frisvad & al. (1990a) considered *E. zonatum* a separate species, characterized by its distinct profile of secondary metabolites and very rough conidia. *Penicillium pulvillorum* var. *echinulatum* may very well represent the anamorph of this species.

Penicillium radiatolobatum Lorinczi in Publ. Soc. Nat. Rom. Pent. Stiinta Sol. 10B: 435. 1972.

Penicillium radiatolobatum is considered to be a synonym of *P. canescens*. Like *P. murcianum* it is a transition form towards *P. janczewskii*.

**Penicillium rubefaciens* Quintanilla in Mycopathologia 80: 73. 1982.

Penicillium rubefaciens resembles *P. raciborskii*, but it has a distinct profile of secondary metabolites.

**Penicillium sajarovii* Quintanilla in Avian. Nutr. Mej. Anim. Alim. 22: 539. 1981.

This species is related to *P. simplicissimum* and *P. canescens*. The growth rate of the ex-type culture is similar to that of *P. simplicissimum*, but the identity has not been confirmed chemically.

Penicillium severskii Schechovtsov in Microbiologia 43: 122. 1981.

Penicillium severskii is a synonym of *P. soppii*, based on the morphology and identical secondary metabolite profiles. It is not a synonym of *P. raciborskii* as stated by Ramírez (1985).

**Penicillium shennongjianum* Kong & Qi in Mycosystema 1: 110. 1988.

The taxon is here accepted, being close to *P. citrinum* and *P. miczynskii*. It differs from these taxa by slow growth on all media (12–16 mm diam. after one week at 25°C), its inabil-

ity to grow on nitrate as sole nitrogen source and to grow at 37°C. The profile of secondary metabolites does not include citrinin and citreoviridin characteristic for *P. citrinum* and *P. miczynskii*, respectively, but other compounds.

****Penicillium smithii*** Quintanilla in *Avan. Nutr. Mej. anim. Alim.* 23: 340. 1982.

Penicillium smithii is here accepted as a distinct species and not a synonym of *P. raciborskii*, as supposed by Ramírez (1985). These two species differ in their growth rates, their ability to produce sclerotia, the roughness of the stipes and profiles of secondary metabolites. The two species have a wide distribution and have been found repeatedly in soil, peat, wood, and on dried fish. All isolates produced great amounts of citreoviridin.

Penicillium turolense Ramírez & Martínez in *Mycopathologia* 74: 36. 1981.

Penicillium turolense is a synonym of *P. westlingii* Zaleski (Frisvad & Filtenborg, 1990). Both species produce large amounts of citrinin.

****Penicillium vasconiae*** Ramírez & Martínez in *Mycopathologia* 72: 189. 1980.

Penicillium vasconiae is a good species, related to *P. daleae* Zaleski, but with conidia lacking transverse ridges.

Penicillium subgenus Penicillium

****Penicillium aethiopicum*** Frisvad apud Frisvad & Filtenborg in *Mycologia* 81: 847. 1989.

Isolates of this species has been identified by other taxonomists as *P. cyclopium*, *P. verrucosum* var. *corymbiferum*, *P. crustosum* or *P. expansum*. Bridge & al. (1989a and b) regarded it as a tropical variant of *P. expansum*, while Pitt & Cruickshank (1990) accepted *P. aethiopicum*. It differs from all the species mentioned above in its growth at 37°C, yellow reverse colours on CYA, MEA and YES, long coherent chains of ellipsoidal conidia, poor growth on creatine-sucrose agar, rough-walled stipes on MEA and the production of griseofulvin and viridicatum-toxin.

Penicillium allii Vincent & Pitt in *Mycologia* 81: 300. 1989.

When describing *P. allii*, Vincent & Pitt (l.c.) regarded it as being closely related to *P. crustosum* and *P. roqueforti*, but they stated that amylase isoenzyme patterns resemble those of *P. hirsutum* and *P. hordei*. This relationship has been further confirmed in that several secondary metabolites in *P. hirsutum* and its varieties are also found in *P. allii* (Frisvad & Filtenborg, 1989). Because of other similarities also (fast growth on most media, rough stipes, smooth globose conidia and association with bulbs and onions) we regard *P. allii* only as a variety of *P. hirsutum* (see also below).

****Penicillium aurantiogriseum*** Dierckx var. *melanoconidium* Frisvad apud Frisvad & Filtenborg in *Mycologia* 81: 848. 1989.

This variety was distinguished from *P. aurantiogriseum* var. *aurantiogriseum* by dark green conidia, a yellow reverse on CYA, rich sporulation on YES and consistent production of

oxaline and penitrem A. Isolates of this variety formed a distinct cluster in the numerical taxonomy of subgenus *Penicillium* by Bridge & al. (1989a).

****Penicillium aurantiogriseum* Dierckx var. *neoechinulatum* Frisvad & al. in Can. J. Bot. 65: 767. 1987.**

The variety *neoechinulatum* is reminiscent of *P. echinulatum* Raper & Thom ex Fassatióvá, but differs from it by smaller blue-green conidia, poor growth on creatine-sucrose agar and production of aurantiamin, penicillic acid and questiomycin. This variety was also considered distinct by Bridge & al. (1989a).

***Penicillium aurantiogriseum* Dierckx var. *polonicum* (Zaleski) Frisvad apud Frisvad & Filtenborg in Mycologia 81: 849. 1989.**

Penicillium polonicum Zaleski was considered to be close to *P. aurantiogriseum* var. *aurantiogriseum*: it only differs from it by the faster growth rates on all media, good sporulation on YES agar, consistent production of penicillic acid and verrucosidin combined with inability to produce xanthomegnin, and terrestric acid. Perhaps only studies on DNA-RNA relationships between the different varieties of *P. aurantiogriseum* can elucidate whether some or all of them should be regarded as species, varieties or chemotypes.

***Penicillium brevicompactum* Dierckx var. *magnum* Ramírez in Man. Atlas Penicillia: 398. 1982.**

In its morphology, growth rates, and profile of secondary metabolites this taxon is indistinguishable from *P. olsonii* Bain. & Sartory.

***Penicillium brunneostoloniferum* Abe ex Ramírez in Man. Atlas Penicillia: 412. 1982.**

Except for its brown conidia, probably caused by a mutation in the biochemical pathway to melanin, this taxon duplicates typical isolates of *P. brevicompactum* in all aspects. *Penicillium brunneostoloniferum*, like *P. brevicompactum* produced brevianamide A, mycophenolic acid and other bluish fluorescent compounds (short-wave UV light).

****Penicillium chrysogenum* Thom var. *dipodomyis* Frisvad & al. in Can. J. Bot. 65: 766. 1987.**

This variety was distinguished from *P. chrysogenum* Westling var. *chrysogenum* by its very dark green conidia, rough conidiophore stipes, production of some unique secondary metabolites and faster growth rate on CYA at 37°C.

****Penicillium confertum* (Frisvad & al.) Frisvad apud Frisvad & Filtenborg in Mycologia 81: 851. 1989.**

This name was introduced to raise the variety *P. glandicola* var. *confertum* Frisvad & al. to specific level (see below).

****Penicillium coprobium* Frisvad apud Frisvad & Filtenborg in Mycologia 81: 851. 1989.**

Penicillium coprobium can be distinguished from *P. coprophilum* by the formation of sclerotia, dark green conidia, green phialides, an entire colony margin on MEA, a pale reverse on CYA and MEA, a black-currant-like aroma and the production of several specific secondary metabolites including patulin. In contrast *P. coprophilum* does not form sclerotia, produces dull green conidia and hyaline phialides, has a lobate colony margin on MEA, a dark brown reverse on CYA and MEA, a herb-like aroma on all substrates and produces other secondary metabolites, including griseofulvin, meleagin and oxaline (Frisvad & Filtenborg, 1989).

****Penicillium coprophilum*** (Berk. & Curtis) Seifert & Samson in Adv. *Penicillium* and *Aspergillus* Syst.: 145. 1986.

In herbarium studies this name was found to be the oldest available for the distinct species so far known as *P. concentricum* Samson, Stolk & Hadlok (1976).

Penicillium cordubense Ramírez & Martínez in Mycopathologia 74: 164. 1981.

Penicillium cordubense is a typical *P. aurantiogriseum* Dierckx var. *aurantiogriseum*, producing viomellein, xanthomegnin and viridicatin.

****Penicillium glandicola*** (Oud.) Seifert & Samson in Adv. *Penicillium* and *Aspergillus* Syst.: 147. 1986.

In herbarium studies this name was found to be the oldest available for the distinct species so far known as *P. granulatum* Bain. (see Seifert & Samson, 1986).

Penicillium glandicola (Oud.) Seifert & Samson var. *confertum* Frisvad & al. in Can. J. Bot. 65: 769. 1987.

This fungus has only been found once. It appears to be more distant from *P. glandicola* than indicated by Frisvad & al. (l.c.). Good growth at 37°C, production of meleagrins and a compound related or similar to asteltoxin, thin sigmoid stipes, and widely divergent phialides indicate that this taxon deserves specific status as *P. confertum* (see above).

****Penicillium glandicola*** (Oud.) Seifert & Samson var. *glaucovenetum* Frisvad apud Frisvad & Filtenborg in Mycologia 81: 854. 1989.

The variety *glaucovenetum* differs from var. *glandicola* by its more discrete synnemata, bluish green conidia, and smooth stipe walls.

Penicillium glandicola (Oud.) Seifert & Samson var. *mononematosum* Frisvad & al. in Can. J. Bot. 65: 767. 1987. (as var. '*mononematosa*').

Since the description of this variety, several new isolates have been obtained of this taxon, including two isolates from salt marsh soil in Egypt. *Penicillium glandicola* var. *mononematosum* is related to *P. lanosum* and *P. chrysogenum*. It differs from these species by its broad rough stipes, its consistently good growth at 37°C, and its production of viriditoxin, isochromantoxin, cyclopaldic acid, verrucologen, fumitremorgin A and C, and occasionally some of the Raistrick phenols (Frisvad, unpubl.). The variety has been raised to specific rank by Frisvad & al. (1989; see also below).

Penicillium granulatum Bain. var. *globosum* Bridge & al. in J. Gen. Microbiol. 135: 2958. 1989.

Distinction of this variety seems to have little justification. In the study by Bridge & al. (1989a) one of the strains of *P. granulatum* (= *P. glandicola*, IMI 297543) appears in both clusters (of *P. granulatum* var. *granulatum* and var. *globosum*). Varieties are supposed to be based on clear-cut, non-overlapping characters (Hawksworth & al., 1983).

**Penicillium griseofulvum* Dierckx var. *dipodomyicola* Frisvad & al. in Can. J. Bot. 65: 767. 1987.

This variety differs consistently from var. *griseofulvum* by rather dark green conidia, a dark brown reverse on CYA, a higher proportion of simpler penicilli, and its inability to produce roquefortine C. The production of roquefortine C in var. *griseofulvum* is consistent.

**Penicillium hirsutum* Dierckx var. *albocoremium* Frisvad apud Frisvad & Filtenborg in Mycologia 81: 855. 1989.

The variety *albocoremium* differs from var. *hirsutum* by the formation of white synnemata and the production of citrinin and meleagrin.

**Penicillium hirsutum* Diercks var. *allii* (Vincent & Pitt) Frisvad apud Frisvad & Filtenborg in Mycologia 81: 855. 1989.

This taxon was described as *P. allii* by Vincent & Pitt (1989). It, however, shares a number of similarities with other varieties of *P. hirsutum* (see above). Results from a numerical study of members of subgenus *Penicillium* also showed *P. hirsutum* var. *allii* to be a distinct taxon (Bridge & al., 1989a), but a possible link to other species was not discussed by these authors.

Penicillium hirsutum Diercks var. *hordei* (Stolk) Frisvad apud Frisvad & Filtenborg in Mycologia 81: 855. 1989.

Penicillium hordei Stolk has many characters in common with *P. hirsutum* var. *hirsutum* (Pitt, 1980; Frisvad & Filtenborg, 1983, 1989; Bridge & al., 1989a), but, like *P. allii* it has some distinctive diagnostic characters too. The most consistent solution is either to accept all the varieties of *P. aurantiogriseum* and *P. hirsutum* as such or to treat them all as species. Until more molecular data on DNA similarities are available we prefer to use the variety level in such cases, so we provisionally accept *P. hirsutum* var. *hordei*.

**Penicillium hirsutum* Diercks var. *venetum* Frisvad apud Frisvad & Filtenborg in Mycologia 81: 855. 1989.

This variety differs from var. *hirsutum* by its dark blue-green conidia, slower growth rate, and production of viridicatin.

Penicillium hispalense Ramírez & Martínez in Mycopathologia 74: 169. 1981.

A dried type of *P. hispalense* was not prepared and the ex-type culture (UFM 5940) is lost (Ramírez, pers. comm.). The 'polyverticillate' structure of the fungus was probably caused

by degeneration. The illustrations and description of *P. hispalense* suggests that this taxon is *P. hirsutum* Dierckx, but the exact status of the species remain in doubt.

Penicillium mali Gorlenko & Novobranova in Mikol. Fitopatol. 17: 464. 1983.

Penicillium mali is now regarded as a synonym of *P. solitum* (Pitt & Cruickshank, 1990; Stolk & al., 1990).

Penicillium mediolanense Dragoni & Cantoni in Ind. Aliment. 155: 281. 1979 (nom. inval., ICBN Art. 36; as '*P. mediolanensis*').

Penicillium mediolanense was described without a Latin diagnosis and designation of holotype material by Dragoni & Cantoni (1979) and Dragoni & Marino (1979). The morphology of this species is identical with *P. verrucosum*. Its synonymy was further supported by the production of ochratoxin A and the restricted growth.

Penicillium melanochlorum (Samson & al.) Frisvad in Adv. Pen. Asp. Syst.: 330. 1985.

This taxon is now regarded a synonym of *P. solitum* (Pitt & Cruickshank, 1990; Stolk & al., 1990).

**Penicillium mononematosum* (Frisvad & al.) Frisvad in Mycologia 81: 856. 1989.

This taxon is discussed above, under *P. glandicola* var. *monematosum*.

Penicillium nordicum Dragoni & Cantoni in Ind. Aliment. 155: 283. 1979 (nom. inval., ICBN Art. 36); ex Ramírez in Adv. *Penicillium* and *Aspergillus* Syst.: 139. 1986.

The type culture produces hyaline and atypically large conidia (comparable to those of *P. commune*) and seems to be a mutant. Because of its growth rates, reverse colours, reaction on creatine-sucrose agar and copious production of ochratoxin A and B, it is allocated to *P. verrucosum*.

Penicillium olivicolor Pitt in Gen. *Penicillium*: 368. 1980.

Penicillium olivicolor was introduced by Pitt (l.c.) as a name change for *P. ochraceum* Bain. apud Thom, because the latter name had already been used for *P. ochraceum* (Corda) Biourge, *P. ochraceum* (Boudier) Biourge, and *P. ochraceum* Raillo. Apart from its inability to produce green melanin complexes, the type isolate is an atypical *P. viridicatum* Westling (Pitt & Cruickshank, 1990).

**Penicillium roqueforti* Thom var. *carneum* Frisvad apud Frisvad & Filtenborg in Mycologia 81: 857. 1989.

The variety *carneum* differs from var. *roqueforti* by its dark blue-green conidia, pale reverse on all substrates, and production of patulin. It never produces PR-toxin as var. *roqueforti* does.

Penicillium solitum Westling var. *crustosum* (Thom) Bridge & al. in J. gen. Microbiol. 135: 2957. 1989.

Even though *P. solitum* Westling and *P. crustosum* Thom have some characters in common such as the ability to produce a restricted rot in apples, cyclopenin production and rough conidiophore stipes, the differences are very significant. *Penicillium solitum* produces compactin and related compounds, while *P. crustosum* produces penitrem A, roquefortine C, and terrestric acid. Furthermore *P. solitum* grows more slowly, has hydrophilic dark green conidia and does not form conidial crusts. *Penicillium crustosum* grows fast, produces grey-green highly hydrophobic conidia, and typical conidial crusts. With Pitt & Cruickshank (1990) and Stolk & al. (1990) we consider the two species as distinct species (also compare Frisvad & al., 1990c).

****Penicillium vulpinum*** (Cooke & Masee) Seifert & Samson in Adv. *Penicillium* and *Aspergillus* Syst.: 144. 1986.

In herbarium studies this name was found to be the oldest available for a distinct species so far known as *P. claviforme* Bain. (Seifert & Samson, 1986).

Penicillium subgenus Biverticillium

Penicillium aurantioflammiferum Ramírez & al. in Mycopathologia 72: 28. 1980.

This species is in all respects a typical *P. islandicum* Sopp.

****Penicillium coalescens*** Quintanilla in Mycopathologia 84: 115. 1983.

This species resembles *P. dendriticum* Pitt and *P. pseudostromaticum* Hodges & al., but, based on differences in conidial shape, growth rates and colony colours, the species is distinct (see also Samson & al., 1989).

****Penicillium dendriticum*** Pitt in Gen. *Penicillium*: 413. 1980.

This species is distinct both morphologically and chemically (Samson & al., 1989).

****Penicillium eberhardtii*** Yokoyama apud Kobayashi & Yokoyama in Bull. natn. Sci. Mus., Tokyo, Ser. B, 7: 20. 1981 (nom. inval., ICBN Art 36).

This name was introduced without Latin description and based on cultural studies of isolates obtained from immature tissues of *Dendrosphaera eberhardtii* Pat. The conidiophores are described and illustrated as biverticillate penicilli and therefore this anamorph should be placed in subgenus *Biverticillium*. We have not examined the isolates and a more detailed examination is required to identify its correct taxonomic status.

****Penicillium erythromellis*** Hocking apud Pitt in Gen. *Penicillium*: 459. 1980.

This is a distinct species producing great amounts of carbohydrate and red exudate droplets.

Penicillium gaditanum Ramírez & Martínez in Mycopathologia 74: 165. 1981.

Penicillium gaditanum is a synonym of *P. minioluteum* (van Reenen-Hoekstra & al., 1990).

Penicillium ilderdanum Ramírez & al. in *Mycopathologia* 72: 32. 1980.

Because of its good growth at 37°C (better than at 25°C), characteristic conical conidial heads, profile of secondary metabolites, vesiculate stipes and metulae and conidial form, this species is inseparable from *P. piceum* Raper & Fennell.

**Penicillium loliense* Pitt in *Gen. Penicillium*: 450. (1980).

Penicillium loliense resembles *P. proteolyticum* Kamyschko, but differs from it by more roughened conidia and slower growth rate at 37°C.

**Penicillium oblatum* Pitt & Hocking in *Mycologia* 77: 819 (1985).

Penicillium oblatum is a good species, characterized by simple to two- stage-branched penicilli of the *Biverticillium* type and acerosc phialides and therefore, in contrast with Pitt & Hocking (1985), we accommodate it in subgenus *Biverticillium*.

**Penicillium palmae* Samson & al. in *Stud. Mycol.* 31: 135 (1989).

This species is very distinctive. It is somewhat related to *P. isariiforme* Stolk & Meyer, but the latter grows much faster and has longer synnemata. Furthermore, *P. palmae* produces mitorubins, while *P. isariiforme* produces secalonic acid D and citreoviridin (Samson & al., 1989).

**Penicillium panamense* Samson & al. in *Stud. Mycol.* 31: 136 (1989).

Penicillium panamense is characterized by conspicuous synnemata in yellow and orange colours, apiculate conidia, and a strongly coloured basal mycelium (red and yellow). These characters set it apart from *P. vulpinum* (Cooke & Masee) Seifert & Samson, to which isolates of *P. panamense* were first allocated. Both species are strictly synnematosous and do not produce mononematous conidiophores in culture.

**Penicillium pittii* Quintanilla in *Mycopathologia* 91: 75 (1985).

This taxon resembles *P. rubrum* Stoll and *P. minioluteum* sensu Pitt, and a more detailed study is needed to elucidate its taxonomic position.

**Penicillium primulinum* Pitt in *Gen. Penicillium*: 455 (1980).

Penicillium primulinum was introduced for *P. diversum* Raper & Fennell var. *aureum* Raper & Fennell, especially because its very characteristic arrangement of the metulae. A second isolate, included in this taxon by Pitt (1980), ATCC 24100, is however a typical *P. marneffei* Segretain (Samson & Frisvad, in prep.).

**Penicillium rademicii* Quintanilla in *Mycopathologia* 91: 72 (1985).

By its poor growth on Czapek agar and its morphology and growth rates this taxon is reminiscent of *P. diversum*, but this identity could not be confirmed by the profiles of secondary metabolites in the two species (van Reenen-Hoekstra & al., 1990).

Penicillium resinae Qi & Kong in *Acta mycol. Sin.* 1: 103. 1982.

This is a synonym of *P. aspersorum* G. Smith and duplicates the latter species in all respects.

****Penicillium sabulosum*** Pitt & Hocking in *Mycologia* 77: 818 (1985).

This is a good species and resembles *P. diversum* Raper & Fennell and *P. tardum* Thom. Because of the unique combination of characters, this species should also be keyed out in subgenus *Furcatum*.

Penicillium samsonii Quintanilla in *Mycopathologia* 91: 69 (1985).

Penicillium samsonii is a synonym of *P. minioluteum* Dierckx (van Reenen- Hoekstra & al., 1990).

****Penicillium siamense*** Manoch & Ramírez in *Mycopathologia* 101: 32 (1988).

Penicillium siamense appears to be a good species, but it has metabolites in common with *P. diversum*. It differs from this species by its better growth on all media.

Penicillium zacynthae Ramírez & Martínez in *Mycopathologia* 74: 167 (1981).

This species is considered to be a synonym of *P. allahabadense* Mehrotra & Kumar (van Reenen-Hoekstra & al., 1990).

Paecilomyces pascuus Pitt & Hocking in *Mycologia* 77: 822. 1985; as '*P. pascua*').

Paecilomyces pascuus belongs to *Penicillium* subgenus *Biverticillium* because of its penicillus structure and the phialide shape. Consequently, we propose the combination: *Penicillium pascuum* (Pitt & Hocking) Frisvad, Stolk & Samson, *comb. nov.* for it.

Penicillium pascuum resembles *P. dendriticum* Pitt, but we have not observed synnema production even after exposure to light, and isolates of *P. pascuum* produce only a few, if any, red or yellow pigments, in contrast to *P. dendriticum*.

Penicillium* subgenus *Geosmithia

Penicillium turris-painense Ramírez in *Mycopathologia* 91: 93 (1985) (as '*P. turris-painensis*').

This taxon is indistinguishable from *P. namyslowskii* Zaleski in all respects.

Geosmithia viridis Pitt & Hocking in *Mycologia* 77: 822 (1985) (as '*G. virida*').

Stolk & Samson (1985) did not recognize *Geosmithia* Pitt as a genus, but considered it as a subgenus of *Penicillium*, because it is difficult to use the morphological characters of species belonging to this group to separate it from the other subgenera with the variable structures e.g. monoverticillate versus biverticillate penicilli, flask-shaped versus acerose phialides, *Eupenicillium* versus *Talaromyces* teleomorphs. If *Geosmithia* was accepted, then separate genera for the subgenera *Biverticillium* and *Aspergilloides* should also be proposed.

Since we reject *Geosmithia* as genus, we propose *Penicillium viride* (Pitt & Hocking) Frisvad, Samson & Stolk, *comb. nov.* *Penicillium viride* produces a diffusible red pigment in MEA after prolonged incubation at low temperatures, a character not observed in other species in subgenus *Geosmithia*.

Table III. List of *Penicillium* nomina nuda which appeared in collection catalogues and patents since 1977.

Nomina nuda	Culture number	Identity (mycotoxins)
<i>P. alloreensis</i> Swanson	ATCC 20399	<i>P. rugulosum</i> Thom (rugulosin)
<i>P. barcinonense</i> Ramírez & Martínez	CBS 330.79	<i>P. corylophilum</i> Dierckx
<i>P. betaolens</i> Ramírez & Martínez	CBS 331.79	<i>P. simplicissimum</i> (= <i>P. janthinellum</i>)
<i>P. citrinum</i> var. <i>pseudopaxilli</i> Martínez & Ramírez	CBS 688.77	<i>P. citrinum</i> Thom, chemotype II (citri- nin and terrein)
<i>P. fungisticum</i> P. C. Misra	ATCC 18089	<i>P. capsulatum</i> Raper & Fennell
<i>P. glaucocoeruleum</i> Ferrer-Ortega & Ramírez	CBS 692.77	<i>P. aurantiogriseum</i> (penicillic acid, ver- rucosidin, cyclopenin)
<i>P. janthinellum</i> var. <i>kuensanii</i> Kinoshita & al.	ATCC 13154	<i>P. simplicissimum</i>
<i>P. mariaecrucis</i> var. <i>fulvescens</i> Quintanilla	ATCC 48476	<i>P. mariaecrucis</i> (xanthomegnin, vio- mellein)
<i>P. ochraceoviride</i> Ferrer-Ortega & Ramírez	CBS 690.77	<i>P. aurantiogriseum</i> (penicillic acid)
<i>P. pimprinum</i> A. Subramanian & Thirumalachar	CBS 373.75	<i>T. emersonii</i> Stolk
<i>P. pinsporum</i> Ramírez & Martínez	IMI 265388A	<i>P. rugulosum</i> (rugulosin)
<i>P. piperis</i> Ramírez & Gonzales	CBS 406.73	<i>P. argillaceum</i> Stolk & al.
<i>P. poonense</i> A. Subramanian & Thirumalachar	CBS 204.75	<i>T. emersonii</i>
<i>P. rinesinum</i> Swanson	ATCC 20398	<i>P. variabile</i> Sopp (rugulosin)
<i>P. restrictum</i> var. <i>kuensanii</i> Kinoshita & al.	ATCC 13155	<i>P. restrictum</i>
<i>P. verrucosum</i> var. <i>cyclopium</i> strain <i>ananas-olens</i>	IJFM 3865	<i>P. chrysogenum</i> Thom

GENUS TALAROMYCES

**Talaromyces assiutensis* Samson & Abdel-Fattah in *Persoonia* 9: 501. 1978

Anamorph: *Penicillium assiutense* Samson & Abdel-Fattah.

This is a distinct species, described and illustrated by Samson & Abdel-Fattah (1978).

**Talaromyces derxii* Takada & Udagawa in *Mycotaxon* 31: 418. 1988.

Anamorph: *Penicillium derxii* Takada & Udagawa.

This distinct heterothallic species resembles *T. bacillisporus* Swift, by its dark green reverse, but differs from it by echinulate ascospores and the anamorph. Until now it is the only heterothallic teleomorph with a *Penicillium* anamorph.

Talaromyces gossypii Pitt in *Gen. Penicillium*: 500. 1980.

Anamorph: *Penicillium gossypii* Pitt.

This species is inseparable from *T. assiutensis* (Frisvad & al., 1990b).

**Talaromyces macrosporus* (Stolk & Samson) Frisvad & al. in *Antonie van Leeuwenhoek* 57: 186. 1990.

Anamorph: *Penicillium macrosporum* Frisvad & al.

The species was introduced to raise *T. flavus* (Kloecker) Stolk & Samson var. *macrosporus* Stolk & Samson to specific rank, because of the distinct profile of secondary meta-

bolites and larger ascospores which possess a higher heat-resistance (see Frisvad & al., 1990b).

**Talaromyces mimosinus* Hocking apud Pitt in *Gen. Penicillium*: 507. 1980.

Talaromyces mimosinus is a distinct species because of its ascospores which are ornamented with distinct sinuous flanges.

**Penicillium vonarxii* Frisvad & Samson in *Antonie van Leeuwenhoek* 57: 186. 1990.

This name was proposed because no name was available for the anamorph of *T. luteus* (Zukal) C.R. Benjamin as *P. luteum* Zukal was described inclusive of the teleomorph.

FURTHER NOMINA NUDA IN *PENICILLIUM*

A number of epithets appear in culture collection catalogues and patents and to our knowledge they have never been validly published. We have reidentified most of these isolates and they are listed in Table III. They produce many yet unidentified secondary metabolites and some of these secondary metabolites may have interesting biotechnological applications.

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